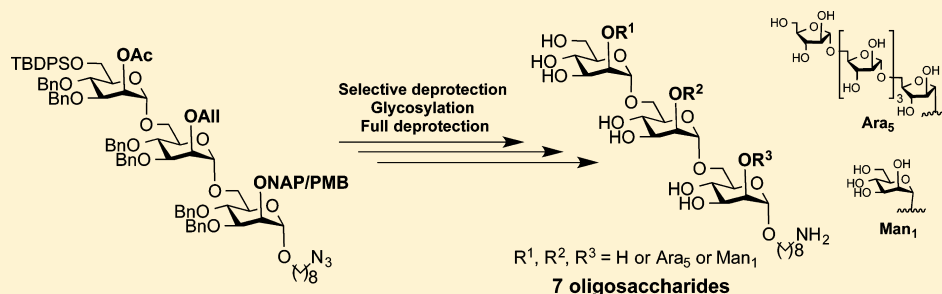


Development of an Orthogonal Protection Strategy for the Synthesis of Mycobacterial Arabinomannan Fragments

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S Supporting Information



ABSTRACT: *Mycobacterium tuberculosis*, the organism that causes tuberculosis (TB), has a carbohydrate-rich cell wall structure that possesses a number of immunogenic antigens. Circulating antibodies that recognize these glycans are present in patients infected by mycobacteria; detection of these antibodies could be the basis for new TB diagnostics. We describe here the synthesis of a panel of mycobacterial arabinomannan fragments for use in investigations directed at testing the feasibility of such a diagnostic method. In this study, we focused on structural motifs present in the core of the key immunogenic polysaccharide lipoarabinomannan (LAM). To access these compounds, we developed an efficient orthogonal protection strategy that allowed access to seven arabinomannan fragments of LAM (1–7). The targets included one tetrasaccharide, one pentasaccharide, three octasaccharides, and two nonasaccharides. Starting from a differentially protected trimannopyranoside derivative (8 or 9), the targets were obtained using an approach that involved selective removal of the protecting group present at the O-2 position of a single mannopyranoside residue, followed by glycosylation with a pentaarabinofuranose thioglycoside and/or a mannopyranose trichloroacetimidate.

INTRODUCTION

Tuberculosis (TB) continues to be one of the most important infectious diseases and leading causes of death worldwide. In 2013, 1.5 million people died from TB and another 9 million fell ill with the disease.¹ Access to reliable and cost-effective diagnostics for infections caused by *Mycobacterium tuberculosis* (the causative agent of TB) and other mycobacteria remains a challenge and is an area of significant research interest.² Like all mycobacteria, *M. tuberculosis* possesses a carbohydrate-rich cell wall that protects the organism from the environment and influences the host immune response upon infection.³ For example, previous work has shown that these glycans modulate cytokine induction⁴ and also lead to a robust antibody response in the host.^{3b,5} Thus, detecting the presence of antibodies that recognize various mycobacterial cell wall glycans could potentially be used in the diagnosis of TB.

A major mycobacterial cell wall glycan is lipoarabinomannan (LAM), a lipidated polysaccharide that plays a critical role in mycobacteria–host interactions.⁶ The interaction of the host with this polysaccharide leads to significant titers of anti-LAM antibodies, underscoring the potential of LAM-based serology in TB diagnosis. Indeed, in a previous study, we demonstrated that detecting antibodies against a hexasaccharide fragment of LAM could discriminate between TB and non-TB patients.⁷

The assay relied on the use of a synthetic derivative of this hexasaccharide, which was immobilized and used in ELISA. The sensitivity and specificity of this diagnostic were enhanced by inclusion of antibody responses against two protein antigens. We hypothesized that antibodies against other domains of LAM could also enhance the performance of the diagnostic and set out to prepare other fragments of this polysaccharide to test this possibility.

The structure of LAM, shown in a schematic form in Figure 1A, consists of a phosphatidyl-*myo*-inositol moiety, a core mannan, an arabinan domain, and a terminal capping motif at the “nonreducing” end of the molecule.^{6,8} The core mannan consists of α -(1→6)-linked D-mannopyranose residues attached to the O-6 position of the inositol. Approximately half of these mannose residues are elaborated with branches consisting of a single α -(1→2)-D-mannopyranose motif. The mannan is further functionalized with an arabinan domain, containing mostly α -(1→5)-linked D-arabinofuranosyl chains with periodic α -(1→3)-linked branch points and terminal β -(1→2)-arabinofuranose residues. These β -linked arabinofuranose residues serve as the site to which a series of capping motifs (e.g., short

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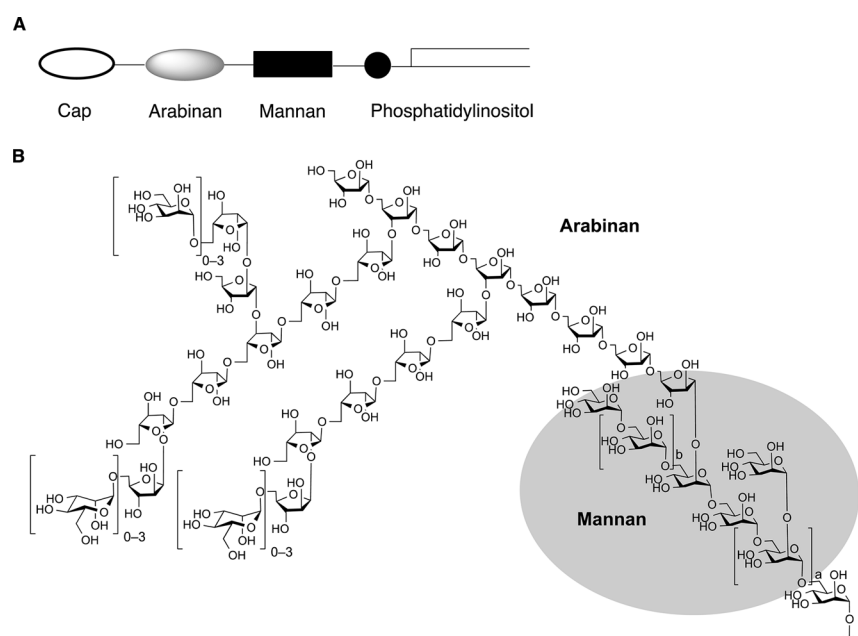


Figure 1. (A) Schematic depiction of LAM. (B) Composite structure of LAM highlighting the arabinomannan domain. The targets prepared in this study correspond to the region shaded in gray.

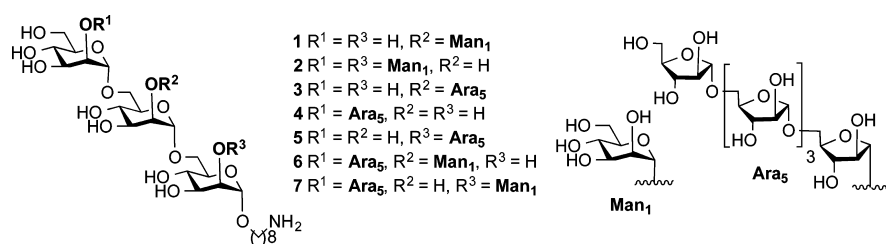
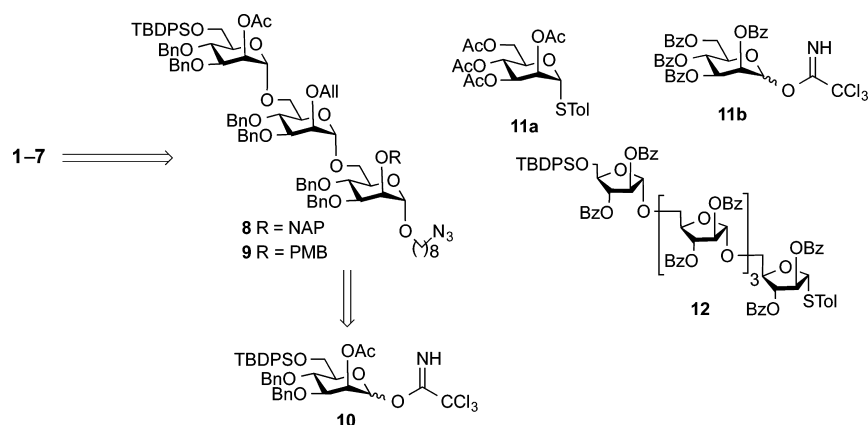


Figure 2. Oligosaccharide targets 1–7.

Scheme 1. Retrosynthetic Analysis of 1–7

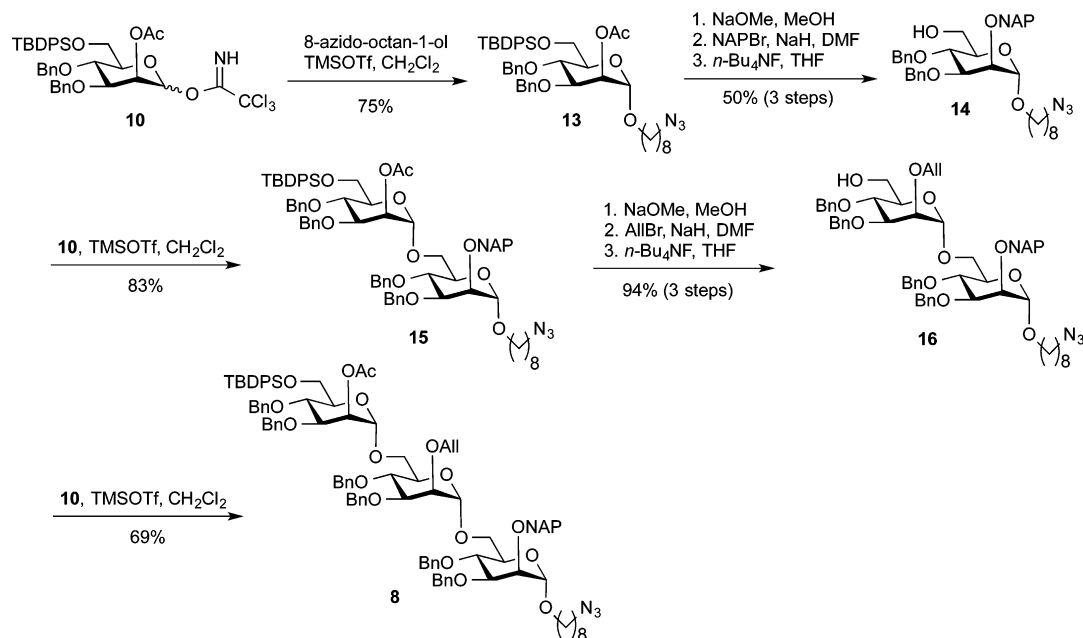


mannopyranosyl oligosaccharides or inositol phosphate moieties) are attached.^{8c,d} A more detailed structure of the mannan core, and its attachment to the arabinan, is shown in Figure 1B.

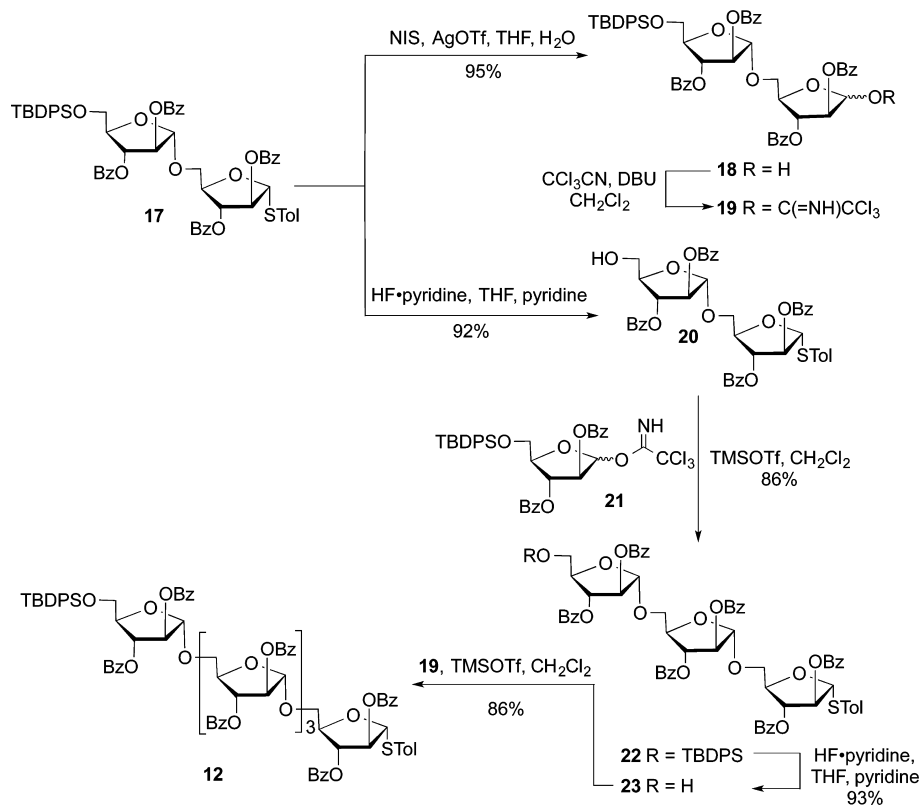
The hexasaccharide used in the aforementioned diagnostic⁷ is a fragment found at the “nonreducing” terminus of the arabinan domain. In choosing additional targets for synthesis, we turned our attention to the mannan core in the hopes of generating structures that would target another subset of anti-LAM antibodies. Thus, we describe here the synthesis of seven oligosaccharides (1–7), which are anticipated fragments of the

core arabinomannan domain of LAM (Figure 2). The oligosaccharides, ranging in size from a tetrasaccharide to a nonasaccharide, include those containing solely the mannan domain (1 and 2), as well as others containing both the mannan and the arabinan domains (3–7). The targets were designed based on the structural motifs suggested to be present in this region of LAM (Figure 1B). In particular, all of the compounds feature an α -(1→6)-mannopyranose backbone, with pendant α -mannopyranose residues and/or an α -(1→5)-linked pentaarabinofuranoside motif attached at O-2 of one of

Scheme 2. Synthesis of Trisaccharide 8



Scheme 3. Synthesis of Pentasaccharide 12

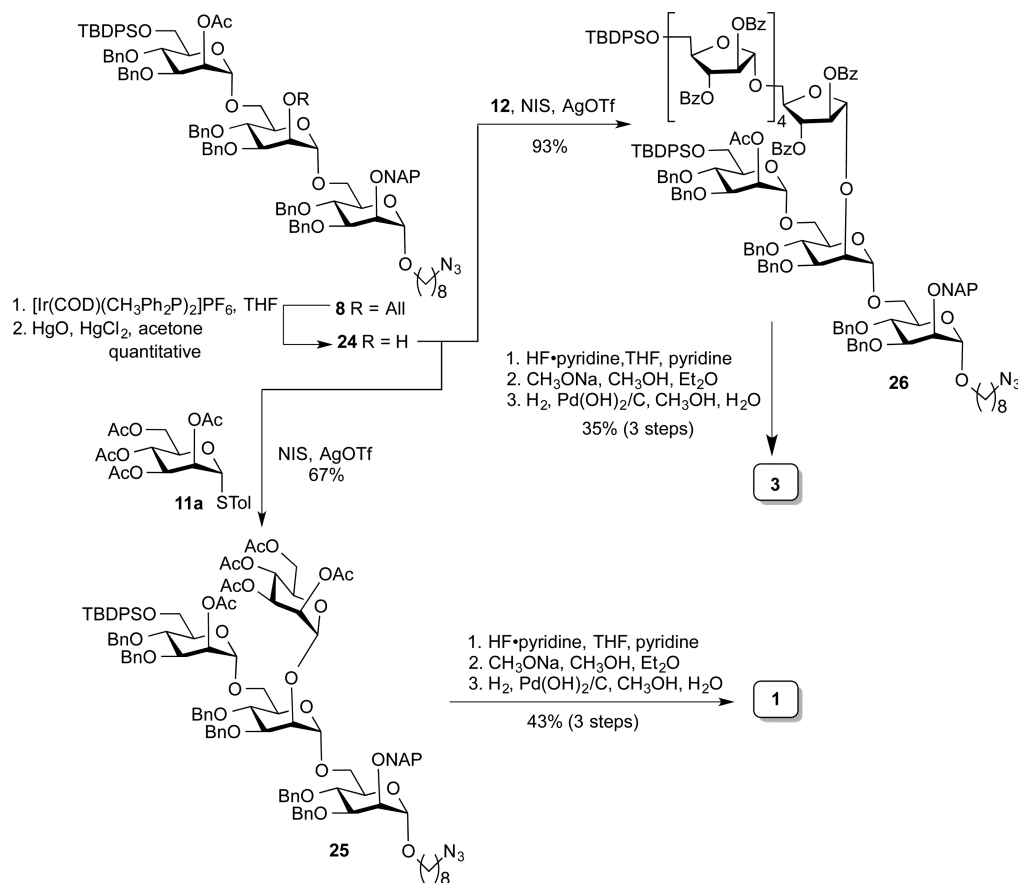


the α -(1 \rightarrow 6)-mannopyranose residues. All oligosaccharides were synthesized with an amino-octyl linker to enable their conjugation to other species (e.g., proteins or solid supports for use in diagnostics). This work complements previous investigations from our group⁹ and others¹⁰ on the synthesis of mycobacterial arabinomannan fragments.

In developing a route to these compounds, we envisioned an orthogonal protection strategy that would allow the preparation of the targets from a common trisaccharide (Scheme 1). One of

the challenges was the need for three orthogonal protecting groups at the O-2 position of each residue in the trimannoside core (8 or 9). The selected protecting groups should be stable to acidic glycosylation conditions, provide α -selectivity to install the required (1 \rightarrow 6) linkages, and have a facile orthogonal deprotection procedure allowing the selective introduction of pentaarabinose (Ara₅) or mannose (Man₁) units as required for the different targets. Ultimately, we relied on a strategy in which the core structure was assembled through the use of glycosyl

Scheme 4. Synthesis of Tetrasaccharide 1 and Octasaccharide 3



donor **10**, with the orthogonal protecting groups being added postglycosylation. The side chain appendages could be added through the use of either monosaccharide donors **11a** and **11b** or pentasaccharide donor **12**.

RESULTS AND DISCUSSION

Implementation of the route outlined in Scheme 1 required access to four building blocks: trisaccharide **8**, monosaccharides **11a** and **11b**, and pentasaccharide **12**. Monosaccharides **11a** and **11b** were prepared as described previously.¹¹ The syntheses of **8** and **12** are described below. In the course of carrying out this work, it was necessary to redesign the trisaccharide building block, and **9** was chosen as a target. The preparation of trisaccharide **9** is described later.

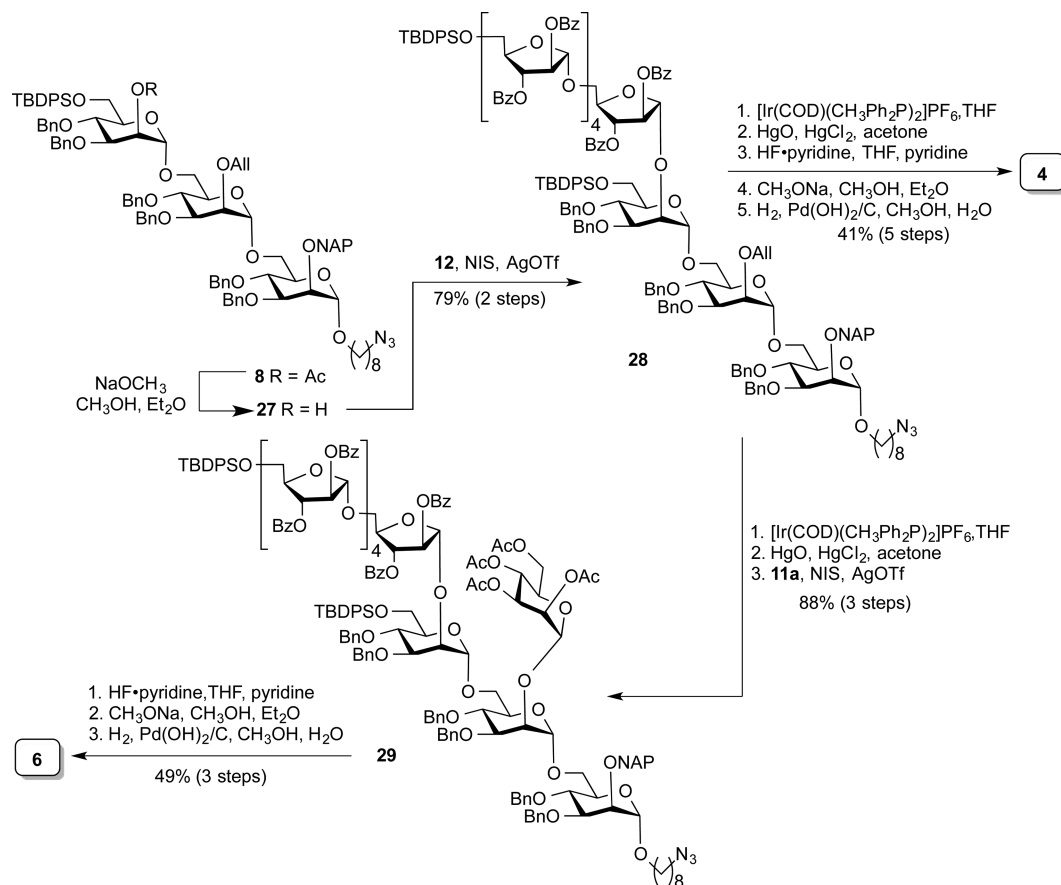
Synthesis of Trisaccharide 8. Trisaccharide **8** was synthesized starting from trichloroacetimidate **10** (Scheme 2).¹² Key features of **10** are the acetyl group on O-2, which secured the required α -selectivity in the glycosylations, and the *tert*-butyldiphenylsilyl (TBDPS) group, which facilitated chain extension. The synthesis began with the glycosylation of 8-azido-octan-1-ol¹³ with **10** activated by trimethylsilyl trifluoromethanesulfonate (TMSOTf), affording **13** in 75% yield. The acetate group was then removed by treatment with sodium methoxide. The resulting hydroxyl group was protected as a naphthylmethyl (NAP) ether, and the TBDPS group was cleaved with tetra-*n*-butylammonium fluoride (*n*-Bu₄NF) in THF, providing alcohol **14** in 50% yield over the three steps. Chain elongation was done by reaction between **14** and **10** using TMSOTf as the promoter to give disaccharide **15** in 83% yield. Disaccharide **16** was obtained in 94% yield following a

similar sequence of deprotection/protection reactions as those described for the preparation of **14**, but introducing an allyl ether instead of a NAP ether. The final mannose residue was added using TMSOTf-promoted glycosylation of **16** with **10**, affording the desired trisaccharide **8** in 69% yield. The α -stereochemistry of the glycosidic linkages in **8** was confirmed via coupled HSQC experiments to measure $^1J_{\text{C-1,H-1}}$ magnitudes. Values of 169, 171, and 172 Hz were obtained, consistent with the α -stereochemistry.¹⁴

Synthesis of Pentasaccharide 12. The synthesis of **12** was carried out starting with disaccharide thioglycoside **17** (Scheme 3), which was prepared as described previously.¹⁵ Cleavage of the silyl ether protecting group in **17** was achieved upon reaction with HF·pyridine to give glycosyl acceptor **20**¹⁶ in 92% yield. Alternatively, the thiotoluy group was hydrolyzed using NIS and AgOTf activation in aqueous THF to afford, in 95% yield, the corresponding lactol **18**, which subsequently was converted into trichloroacetimidate **19**.

Having accessed **19** and **20**, glycosylation of the latter with the trichloroacetimidate derivative **21**¹⁵ afforded **22** in 86% yield. The TBDPS group was then deprotected using HF·pyridine to provide the trisaccharide acceptor **23** in 93% yield. The synthesis of the pentaarabinose building block **12** was then achieved, in 86% yield, through glycosylation of **23** with **19** in the presence of TMSOTf in CH_2Cl_2 . The α -anomeric configuration of the glycosidic linkages in **12** was confirmed from the ¹³C NMR spectrum on the basis of the four signals clustered around 106.0 ppm and a fifth at 91.6 ppm, the latter corresponding to the anomeric carbon of the residue bearing the thiotoluy group.^{9,15,17} Furthermore, all five H-1 signals

Scheme 5. Synthesis of Octasaccharide 4 and Nonasaccharide 6



appear as singlets in the ^1H NMR spectrum, consistent with previous literature for α -arabinofuranosides.^{9,11}

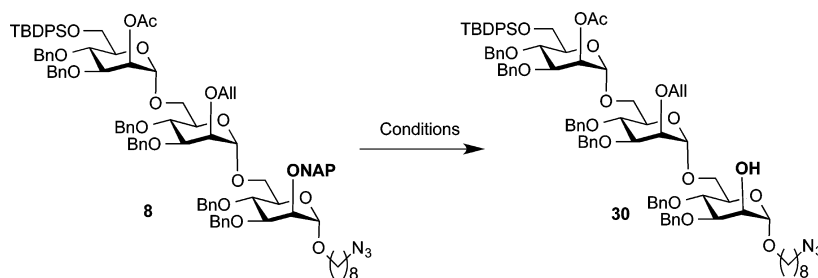
Synthesis of Oligosaccharides (1–7). With trisaccharide 8, monosaccharides 11a and 11b, and pentasaccharide 12 in hand, we next turned our attention to the synthesis of 1–7. To accomplish this goal, selective deprotection of the acetate, allyl, or naphthylmethyl groups present at the O-2 positions of 8 is required. Although previous literature suggested that these selective deprotection steps would be straightforward, as outlined in the discussion below, we faced challenges that needed to be overcome.

Synthesis of Tetrasaccharide 1 and Octasaccharide 3. Initially, we focused on the synthesis of tetrasaccharide 1 and octasaccharide 3, which required the selective cleavage of the allyl group in 8 (Scheme 4). Our first attempt to directly remove the allyl group in 8 using PdCl_2 under buffered conditions ($\text{AcOH}, \text{NaOAc}$)¹⁸ led to decomposition of the starting material. On the other hand, attempted deprotection in the presence of a catalytic amount of $\text{Pd}(\text{PPh}_3)_4$ ¹⁹ did not proceed; only the starting material remained. Therefore, indirect approaches involving allyl group isomerization and then hydrolysis were explored. The use of Wilkinson catalyst²⁰ $\text{RhCl}(\text{Ph}_3\text{P})_3$ gave a poor yield of the vinyl ether isomerization product (<20%). In contrast, the use of $[\text{Ir}(\text{COD})(\text{CH}_3\text{Ph}_2\text{P})_2]\text{PF}_6$ ²¹ resulted in complete conversion of 8 into the corresponding vinyl ether. Subsequent hydrolysis using HgO and HgCl_2 ²² in wet acetone produced the desired alcohol 24 in quantitative yield.

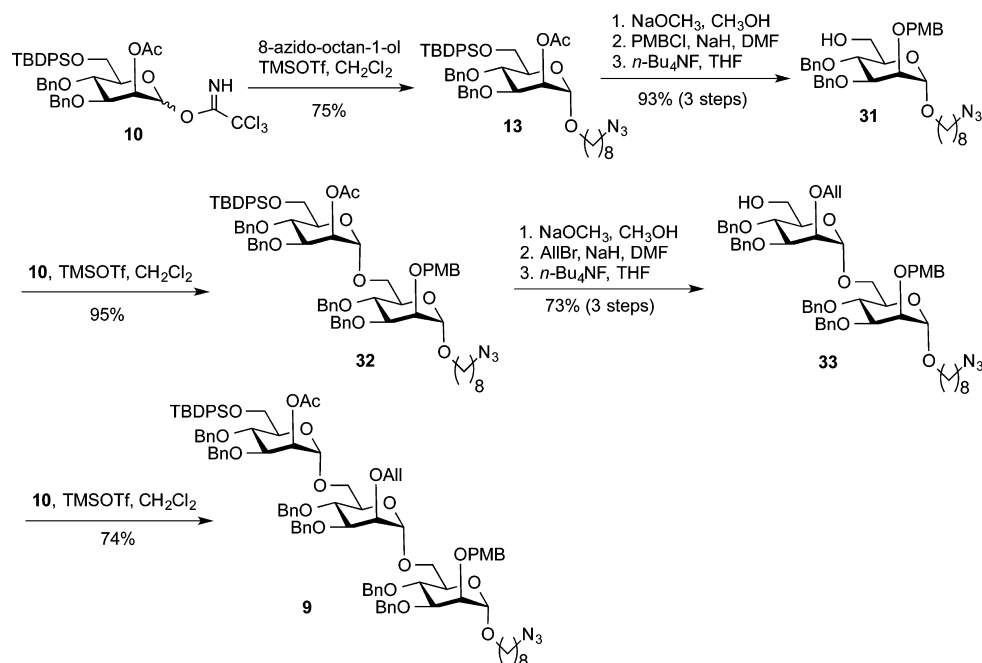
Trisaccharide 24 was further coupled with an excess of 11a^{11a} using NIS–AgOTf activation at 0 °C to afford

tetrasaccharide 25 in 67% yield with complete α -selectivity ($^1J_{\text{C-1,H-1}} = 171$ Hz). In contrast, the glycosylation of 24 with pentasaccharide thioglycoside 12 under the same conditions did not proceed. However, complete conversion of the starting materials was observed when the reaction was carried out at room temperature, leading to the formation of the desired octasaccharide 26 in 93% yield. Tetrasaccharide 1 and octasaccharide 3 were obtained after removal of the protecting groups and conversion of the azide to an amine in a three-step protocol. The silyl ether was cleaved in the presence of $\text{HF}\cdot\text{pyridine}$, and the esters were cleaved using sodium methoxide. Finally, the azide group was reduced and the benzyl protecting groups were removed by hydrogenolysis using $\text{Pd}(\text{OH})_2/\text{C}$ in methanol and water to afford 1 and 3 in 43% and 35% yield, respectively, over three steps.

Synthesis of Octasaccharide 4 and Nonasaccharide 6. To synthesize 4 and 6, an approach similar to that used for the preparation of 1 and 3 was employed, but involving the selective deprotection of the acetate ester in trisaccharide 8 (Scheme 5). Thus, treatment of 8 with sodium methoxide in methanol and dichloromethane afforded 27, which was used without further purification. Glycosylation of 27 with pentasaccharide thioglycoside 12 afforded the fully protected octasaccharide 28 in 79% yield with complete α -selectivity. Once 28 had been obtained, we could synthesize nonasaccharide 29 after selective deprotection of the allyl group using $[\text{Ir}(\text{COD})(\text{CH}_3\text{Ph}_2\text{P})_2]\text{PF}_6$ as described above, followed by glycosylation of the resulting alcohol with thioglycoside donor 11a under NIS–AgOTf activation. Nonasaccharide 29 was obtained in 88% overall yield from 28. Deprotection of the

Table 1. Attempted Cleavage of NAP Ether in **8**

entry	conditions	result
1	DDQ (4 equiv), CH ₂ Cl ₂ , CH ₃ OH or H ₂ O, rt.	low yield + debenzoylation
2	DDQ (2 equiv), CH ₂ Cl ₂ /MeOH, 0 °C	low conversion + debenzoylation
3	DDQ (10%), FeCl ₃ , CH ₂ Cl ₂ /H ₂ O, rt.	no reaction
4	CAN (2–6 equiv), CH ₃ CN/H ₂ O, rt.	low conversion + debenzoylation
5	HF·pyridine, toluene, rt.	low conversion + desilylated product

Scheme 6. Synthesis of Trisaccharide **9**

TBDPS ether, benzoate esters, benzyl ethers and azide reduction was carried out as described for **1** and **3** to provide octasaccharide **4** and nonasaccharide **6** in 41% and 49% yield, respectively.

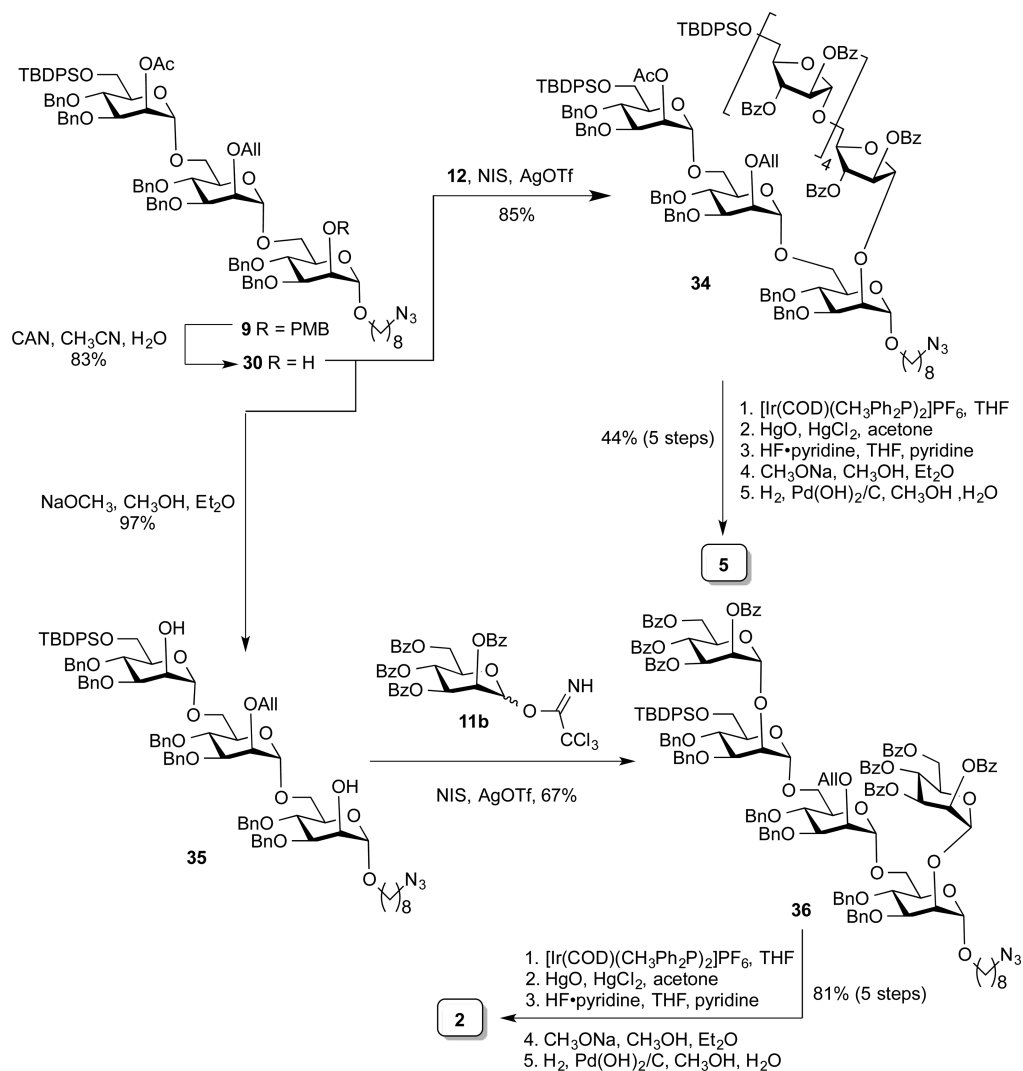
Synthesis of Pentasaccharide **2** and Octasaccharide **5**.

To synthesize oligosaccharide targets **2** and **5**, the selective deprotection of the NAP ether in trisaccharide **8** was required. Our first attempts made use of oxidative procedures (Table 1, entries 1–4). When **8** was treated with 4 equiv of 2,3-dichloro-5,6-dicyano-*p*-benzoquinone (DDQ)²³ in dichloromethane with methanol or water at room temperature, a mixture of several compounds was obtained, presumably due to the deprotection of the benzyl groups as well as cleavage of the NAP ether. Reducing the amount of DDQ to 2 equiv and using lower reaction temperatures (0 °C) did not improve the selectivity. The use of a catalytic amount of DDQ²⁴ was also explored, but the reaction did not proceed. Another oxidative agent, ceric ammonium nitrate (CAN),²⁵ was also investigated. Using 2 equiv of the oxidant, a low (<10%) conversion into the

desired product was observed. Increasing the amount of CAN to 6 equiv led to a mixture of compounds. Given the failure of oxidative methods to affect the cleavage of the NAP ether, we turned to a recently reported method described by Liu and co-workers, which makes use of HF·pyridine in toluene (Table 1, entry 5).²⁶ However, although this method has been shown to succeed with molecules containing silyl protecting groups, applying these conditions to **8** did not lead to the cleavage of the NAP ether. Instead, only the desilylation product was isolated. The difficulties encountered in the selective cleavage of the NAP ether in **8** mirror other (unpublished) results from our laboratory. We have often found it difficult to cleave selectively NAP ethers in the presence of large numbers of benzyl ethers (here six), without significant amounts of decomposition, which we assume is competitive debenzoylation.

Disappointed by these results, we altered the strategy and replaced the NAP ether with a *p*-methoxybenzyl (PMB) ether, which we anticipated could be removed selectively in the presence of acetate esters, allyl ethers, TBDPS ethers, and

Scheme 7. Synthesis of Pentasaccharide 2 and Octasaccharide 5



benzyl ethers. Thus, we synthesized trisaccharide **9**, which differs from **8** by the substitution of the NAP ether with a PMB ether. As outlined in Scheme 6, compound **9** could be obtained in nine steps starting from the known trichloroacetimidate derivative **10** in 36% overall yield using the same route as that described above for trisaccharide **8**.

Once compound **9** was available, the selective deprotection of the PMB ether was investigated. We found that the use of CAN²⁷ in acetonitrile and water cleanly provided the desired alcohol **30** in 83% yield (Scheme 7). Having successfully removed the PMB ether, acceptor **30** was coupled with the pentasaccharide thioglycoside **12** under NIS and AgOTf activation. The expected octasaccharide, **34**, was isolated in 85% yield. Deprotection of the allyl group was achieved upon treatment with [Ir(COD)(CH₃Ph₂P)₂]PF₆, followed by hydrolysis of the resulting vinyl ether using HgO and HgCl₂. Subsequent deprotection of the silyl ether and ester groups and then hydrogenolysis provided the desired octasaccharide **5** in 44% yield over the five steps. Alternatively, methanolysis of the acetate ester in trisaccharide **30** (yielding diol **35**), followed by glycosylation with trichloroacetimidate derivative **11b** under TMSOTf activation, afforded pentasaccharide **36** in 67% yield. We note that the use of thioglycoside **11a** for this reaction led to a complex mixture of products. Compound **36** was

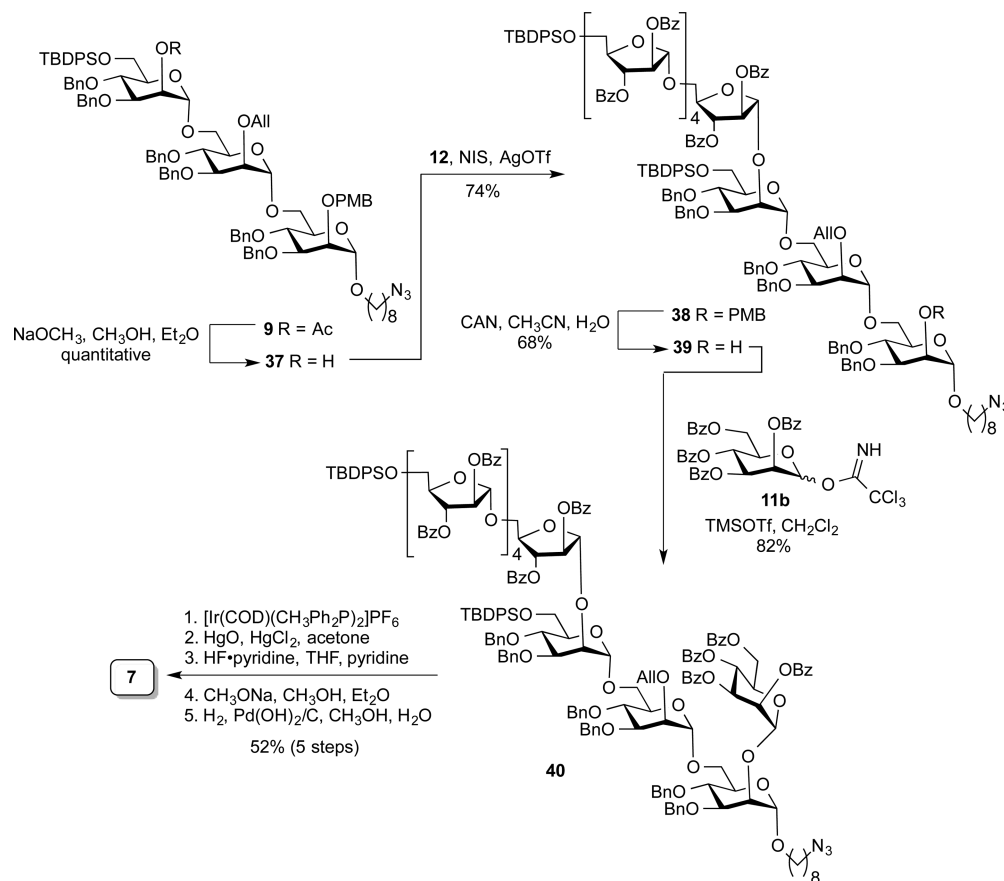
deprotected using the same series of transformations as those described for the other oligosaccharides, giving pentasaccharide **2** in 81% yield over five steps.

Synthesis of Nonasaccharide 7. The final target, nonasaccharide **7**, was prepared as shown in Scheme 8, starting with trisaccharide **9**. Methanolysis of the acetate ester led quantitatively to **37**, which was subsequently coupled to thioglycoside **12**, providing octasaccharide **38** in 74% yield. Subsequently, the PMB group in **38** was cleaved using CAN in acetonitrile and water to give alcohol **39** in 68% yield. Glycosylation of **39** with trichloroacetimidate **11b** led to nonasaccharide **40** in 82% yield. Nonasaccharide **7** was obtained after deprotection of the protecting groups, as carried out for the other targets, in 52% yield over five steps.

CONCLUSION

In summary, we report here an efficient strategy to access seven complex oligosaccharide fragments of the arabinomannan domain of mycobacterial LAM. The route developed was a convergent one requiring as the key intermediate an orthogonally protected trisaccharide derivative (**8** or **9**), which could be selectively deprotected to liberate one of three hydroxyl groups. Initially, we chose the three orthogonal protecting groups to be an acetate ester, allyl ether, and NAP

Scheme 8. Synthesis of Nonasaccharide 7



ether (trisaccharide **8**). However, difficulties in the selective cleavage of the NAP ether led us to develop a different intermediate in which this group was replaced with a PMB ether (trisaccharide **9**). This underscores a problem we have frequently encountered in the synthesis of other complex glycans (to be reported elsewhere) where late stage cleavage of a NAP ether has been problematic and has forced the redesign of a route. Overall, we find the selective removal of a NAP ether in the presence of a large number of benzyl groups to be unpredictable, and its use in complex glycan synthesis should, therefore, be carefully considered. Subsequent glycosylation of the alcohol obtained from **8** or **9** with monosaccharide (**11a/11b**) and/or pentasaccharide (**12**) glycosyl donors gave compounds ranging in size from tetrasaccharide to nonasaccharides, which were then fully deprotected and isolated in good yields. All oligosaccharides were synthesized containing an amino group to facilitate their conjugation to proteins or to potential diagnostic devices. Downstream investigations using these compounds will be reported in the future.

EXPERIMENTAL SECTION

General Methods. All reagents were purchased from commercial sources and were used without further purification unless noted. Reaction solvents were purified by successive passage through columns of alumina and copper under argon. Unless stated otherwise, all reactions were carried out at room temperature and under a positive pressure of argon and were monitored by TLC on Silica Gel G-25 F254 (0.25 mm). TLC spots were detected under UV light and/or by charring with a solution of *p*-anisaldehyde in ethanol, acetic acid, and sulfuric acid. Column chromatography was performed on Silica Gel 60 (40–60 μm). Solvents were evaporated under reduced pressure on a

rotary evaporator. Optical rotations were measured in a microcell (10 cm, 1 mL) at ambient temperature and are in units of degree-mL/(g-dm). ^1H NMR spectra were recorded at 400, 500, 600, or 700 MHz, and chemical shifts are referenced to residual CHCl_3 (7.26 ppm, CDCl_3), HOD (4.78 ppm, D_2O), or CHD_2OD (3.30 ppm, CD_3OD). ^{13}C NMR spectra were recorded at 126, 151, or 176 MHz, and chemical shifts are referenced to CDCl_3 (77.0 ppm) or CD_3OD (48.9 ppm, CD_3OD). Reported splitting patterns are abbreviated as *s* = singlet, *d* = doublet, *t* = triplet, *m* = multiplet, *br* = broad, *app* = apparent. Assignments of NMR spectra were based on two-dimensional experiments (^1H – ^1H COSY, HSQC, and HMBC). High-resolution ESI-MS spectra (time-of-flight analyzer) were recorded on samples suspended in THF or CH_3OH and with added NaCl.

8-Aminoocetyl α -D-Mannopyranosyl-(1 \rightarrow 6)-[α -D-mannopyranosyl-(1 \rightarrow 2)]- α -D-mannopyranosyl-(1 \rightarrow 6)- α -D-mannopyranoside (1**).** To a solution of tetrasaccharide **25** (77 mg, 0.039 mmol) in THF–pyridine (5:2, 700 μL) at 0 $^\circ\text{C}$ was added a solution of HF–pyridine 70% in pyridine (25 μL) dropwise. The reaction mixture was warmed to room temperature and stirred for 2 days. After dilution with EtOAc (10 mL), the reaction mixture was poured into a satd aq solution of NaHCO_3 (15 mL) and extracted with EtOAc (2 \times 10 mL). The organic layer was washed with H_2O (2 \times 10 mL), dried (Na_2SO_4), filtered, and concentrated under vacuum to give a syrup that was filtered through a short silica gel column (4:1 hexane–EtOAc). The residue obtained after solvent evaporation was dissolved in Et_2O – CH_3OH (1:1, 2 mL), before adding a solution of NaOCH_3 in CH_3OH (1 mL, 0.1M). The reaction mixture was stirred overnight, neutralized by the addition of Amberlyst IR-120 (H^+) cation exchange resin, filtered, and concentrated to give a syrup that was used without any further purification. This crude product was dissolved in H_2O – CH_3OH (1:1, 3 mL); then $\text{Pd}(\text{OH})_2/\text{C}$ (10%) was added and the reaction mixture was stirred vigorously under a hydrogen atmosphere (1 atm) overnight. The reaction mixture was diluted with H_2O –

evaporation, the residue was dissolved in water and then lyophilized to give **6** (11.9 mg, 49% over 3 steps) as an amorphous fluffy white solid. $[\alpha]_D + 57.1$ ($c = 0.09$, CH_2Cl_2); $^1\text{H NMR}$ (600 MHz, D_2O) $\delta_{\text{H}} = 5.15$ (br s, 1 H, H-1), 5.10 (br s, 1 H, H-1), 5.08–5.04 (m, 4 H, 4 × H-1), 5.00 (br s, 1 H, H-1), 4.98 (br s, 1 H, H-1), 4.82 (br s, 1 H, H-1), 4.21–4.15 (m, 5 H), 4.09 (br s, 3 H), 4.08–4.03 (m, 2 H), 4.00–3.95 (m, 6 H), 3.94–3.83 (m, 11 H), 3.81–3.58 (m, 21 H), 3.57–3.41 (m, 3 H), 2.94 (t, 2 H, $J = 7.4$ Hz, CH_2NH_2), 1.71–1.55 (m, 4 H, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{NH}_2$), 1.38–1.22 (m, 8 H, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{NH}_2$); $^{13}\text{C NMR}$ (126 MHz, D_2O) $\delta_{\text{C}} = 109.5$ (C-1), 107.5 (2 × C-1), 107.4 (2 × C-1), 102.3 (C-1), 99.9 (C-1), 98.8 (C-1), 98.2 (C-1), 84.0 (2 C), 82.4 (3 C), 82.0, 81.2, 80.9, 80.8 (2 C), 78.8, 77.5, 76.8 (2 C), 76.7 (2 C), 73.4, 73.3, 72.8, 71.2, 71.1, 71.0, 70.5 (2 C), 70.4, 70.1 (3 C), 68.1, 66.9, 66.8 (2 C), 66.7, 66.1, 65.8, 62.8, 62.5, 61.2 (2 C), 61.1, 60.8, 39.6, 28.5, 28.2 (2 C), 26.9, 25.6, 25.3. HRMS (ESI) calcd for ($\text{M} + \text{H}^+$) $\text{C}_{57}\text{H}_{100}\text{NO}_{41}$: 1454.5765. Found: 1454.5761.

8-Amino-octyl α -D-Arabinofuranosyl-(1→5)- α -D-arabinofuranosyl-(1→5)- α -D-arabinofuranosyl-(1→5)- α -D-arabinofuranosyl-(1→5)- α -D-mannopyranosyl-(1→2)- α -D-mannopyranosyl-(1→6)- α -D-mannopyranosyl-(1→6)-[α -D-mannopyranosyl-(1→2)]- α -D-mannopyranoside (7**).** The synthesis of **7** was achieved starting from the nonasaccharide **40** (53 mg, 0.013 mmol) following the procedure described for the compound **2**. The product was purified by gel filtration chromatography (Sephadex, LH-20) using CH_3OH as the eluent. After solvent evaporation, the residue was dissolved in water and then lyophilized to give **7** (10 mg, 52% over 5 steps) as an amorphous fluffy white solid. $[\alpha]_D - 5.4$ ($c = 0.10$, CH_2Cl_2); $^1\text{H NMR}$ (700 MHz, D_2O) $\delta_{\text{H}} = 5.14$ (br s, 1 H, H-1), 5.07–5.02 (m, 5 H, 5 × H-1), 5.00–4.96 (m, 2 H, 2 × H-1), 4.87 (br s, 1 H, H-1), 4.64 (s, 1 H), 4.20–4.14 (m, 4 H), 4.09 (br s, 3 H), 4.07–4.03 (m, 2 H), 3.98–3.88 (m, 11 H), 3.87–3.82 (m, 6 H), 3.81–3.58 (m, 22 H), 3.55–3.47 (m, 2 H), 2.93 (t, 2 H, $J = 7.4$ Hz, CH_2NH_2), 1.67–1.52 (m, 4 H, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{NH}_2$), 1.38–1.26 (m, 8 H, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{NH}_2$); $^{13}\text{C NMR}$ (176 MHz, D_2O) $\delta_{\text{C}} = 109.4$ (C-1), 107.5 (2 × C-1), 107.4 (2 × C-1), 102.4 (C-1), 99.5 (C-1), 98.8 (C-1), 98.2 (C-1), 84.0 (2 C), 82.4 (2 C), 82.3, 82.0, 81.8, 81.1, 80.9 (2 C), 80.8 (2 C), 79.0, 77.5, 76.7 (2 C), 76.5 (2 C), 73.3, 72.7, 71.8, 71.1, 71.0, 70.5 (2 C), 70.3, 70.2, 70.0 (2 C), 68.5, 66.9 (2 C), 66.8 (2 C), 66.5, 65.9, 62.5, 61.2 (2 C), 61.1, 60.8, 39.6, 28.4, 28.2, 28.1, 26.9, 25.5, 25.3; HRMS (ESI) calcd for ($\text{M} + \text{H}^+$) $\text{C}_{57}\text{H}_{100}\text{NO}_{41}$: 1454.5765. Found: 1454.5766.

8-Azido-octyl 2-O-Acetyl-3,4-di-O-benzyl-6-O-(tert-butylidiphenylsilyl)- α -D-mannopyranosyl-(1→6)-2-O-allyl-3,4-di-O-benzyl- α -D-mannopyranosyl-(1→6)-3,4-di-O-benzyl-2-O-(2-methylnaphthyl)- α -D-mannopyranoside (8**).** A mixture of trichloroacetimidate **10** (1.036 g, 1.32 mmol), alcohol **16** (1.103 g, 1.064 mmol), and 4 Å molecular sieves (290 mg) in CH_2Cl_2 (12 mL) was stirred for 30 min at -20 °C under an argon atmosphere. Then, TMSOTf (20 μL , 0.106 mmol) was added dropwise over 5 min. The reaction mixture was warmed to 0 °C over 20 min, and then the TMSOTf was quenched by the addition of Et_3N . The solution was concentrated under vacuum and the resulting syrup was purified by column chromatography (9:1 to 8.5:1.5 hexane–EtOAc) to afford **8** (1.22 g, 69%) as a syrup: R_f 0.59 (4:1 hexane–EtOAc); $[\alpha]_D + 35.3$ ($c = 0.37$, CH_2Cl_2); $^1\text{H NMR}$ (400 MHz, CDCl_3) $\delta_{\text{H}} = 7.83$ –7.64 (m, 8 H, Ar), 7.57–7.08 (m, 39 H, Ar), 5.92 (dddd, 1 H, $J = 5.0, 5.9, 10.4, 17.2$ Hz, $\text{CH}_2\text{-CH=CH}_2$), 5.52 (dd, 1 H, $J = 1.8, 3.4$ Hz, H-2''), 5.30 (dd, 1 H, $J = 1.4, 17.2$ Hz, $\text{CH}_2\text{-CH=CH}_2$), 5.12 (dd, 1 H, $J = 1.4, 10.4$ Hz, $\text{CH}_2\text{-CH=CH}_2$), 5.08 (d, 1 H, $J = 1.3$ Hz, H-1'), 4.97–4.88 (m, 6 H, H-1'', CH_2NAP , 4 × CH_2Ph), 4.87 (d, 1 H, $J = 1.5$ Hz, H-1), 4.72 (d, 1 H, $J = 11.2$ Hz, CH_2Ph), 4.67 (br s, 2 H, CH_2Ph), 4.64–4.57 (m, 2 H, CH_2Ph), 4.56–4.47 (m, 3 H, CH_2Ph), 4.43 (d, 1 H, $J = 11.4$ Hz, CH_2NAP), 4.18–4.05 (m, 3 H, 2 × $\text{CH}_2\text{-CH=CH}_2$, H-3''), 4.05–3.81 (m, 9 H, H-4'', H-2', H-2, H-3, H-3', H-4, H-4', H-6a'', H-6a), 3.80–3.49 (m, 8 H, H-5, H-5', H-5'', H-6b'', H-6a', H-6b, H-6b', OCH_2), 3.34 (dt, 1H, $J = 2 \times 6.4, 9.6$ Hz, OCH_2), 3.23 (t, 2 H, $J = 6.9$ Hz, CH_2N_3), 2.14 (s, 3 H, CH_3), 1.63–1.44 (m, 4 H, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$), 1.41–1.19 (m, 8 H, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$), 1.09 (m, 9 H, $\text{C}(\text{CH}_3)_3$); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) $\delta_{\text{C}} = 170.2$ (C=O), 138.9 (Ar), 138.8 (Ar), 138.6 (Ar), 138.5 (Ar), 138.2 (Ar), 136.0 (2 C, Ar), 135.8 ($\text{CH}_2\text{-CH=CH}_2$), 135.6 (2 C, Ar), 135.2 (Ar), 134.0 (Ar), 133.3 (Ar), 133.2 (Ar), 133.0 (Ar), 129.6 (Ar), 129.5 (Ar), 128.5 (3C, Ar), 128.4 (8 C, Ar), 128.2 (3 C, Ar), 128.1 (2 C, Ar), 128.0 (2 C, Ar), 127.8 (Ar), 127.7 (9 C, Ar), 127.6 (4 C, Ar), 127.5 (2 C, Ar), 127.3 (2 C, Ar), 127.2 (Ar), 126.7 (Ar), 126.2 (Ar), 126.0 (Ar), 125.9 (Ar), 116.9 ($\text{CH}_2\text{-CH=CH}_2$), 98.2 (C-1'), 98.1 (C-1''), 98.0 (C-1), 80.6 (C-3), 79.5 (C-3'), 77.9 (C-3''), 75.1 (2 C, CH_2NAP , CH_2Ph), 75.0 (C-4'), 74.8 (CH_2Ph), 74.7 (2 C, C-4, C-4'), 74.2, 73.9 (C-2, C-2'), 73.1 (CH_2Ph), 72.5 (C-5''), 72.3 (CH_2Ph), 71.5 (CH_2Ph), 71.4 ($\text{CH}_2\text{-CH=CH}_2$), 71.2 (C-5, C-5'), 68.6 (C-2''), 67.6 (CH_2O), 66.2, 66.1 (C-6, C-6'), 62.6 (C-6''), 51.4 (CH_2N_3), 29.4, 29.3, 29.1, 28.8 (4 C, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$), 26.8 (3 C, $\text{C}(\text{CH}_3)_3$), 26.7, 26.1 (2 C, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$), 21.1 (CH_3), 19.4 ($\text{C}(\text{CH}_3)_3$). HRMS (ESI) calcd for ($\text{M} + \text{Na}^+$) $\text{C}_{100}\text{H}_{115}\text{N}_3\text{O}_{17}\text{SiNa}$: 1680.7888. Found: 1680.7880.

8-Azido-octyl 2-O-Acetyl-3,4-di-O-benzyl-6-O-(tert-butylidiphenylsilyl)- α -D-mannopyranosyl-(1→6)-2-O-allyl-3,4-di-O-benzyl- α -D-mannopyranosyl-(1→6)-3,4-di-O-benzyl-2-O-(4-methoxybenzyl)- α -D-mannopyranoside (9**).** The synthesis of **9** was achieved following the procedure described for the compound **8**, using disaccharide **33** (1.3 g, 1.28 mmol) and trichloroacetimidate **10** (1.205 g, 1.53 mmol) in the presence of TMSOTf (35 μL , 0.19 mmol) in Et_2O (16 mL). The crude residue was purified by column chromatography (9:1 to 8:2 hexane–EtOAc) to yield **9** (1.55 g, 74%) as a syrup. R_f 0.45 (4:1 hexane–EtOAc); $[\alpha]_D + 33.5$ ($c = 0.46$, CH_2Cl_2); $^1\text{H NMR}$ (500 MHz, CDCl_3) $\delta_{\text{H}} = 7.76$ (d, 2 H, $J = 6.8$ Hz, Ar), 7.68 (d, 2 H, $J = 6.8$ Hz, Ar), 7.46–7.09 (m, 38 H, Ar), 6.83 (d, 2 H, $J = 8.4$ Hz, PhOCH_3), 5.95 (dddd, 1 H, $J = 5.4, 6.2, 10.5, 17.2$ Hz, $\text{CH}_2\text{-CH=CH}_2$), 5.53 (dd, 1 H, $J = 1.7, 2.9$ Hz, H-2''), 5.34 (dd, 1 H, $J = 1.4, 17.1$ Hz, $\text{CH}_2\text{-CH=CH}_2$), 5.16 (dd, 1 H, $J = 1.4, 10.4$ Hz, $\text{CH}_2\text{-CH=CH}_2$), 5.08 (d, 1 H, $J = 1.2$ Hz, H-1'), 4.95 (d, 1 H, $J = 1.7$ Hz, H-1''), 4.95 (d, 1 H, $J = 10.8$ Hz, CH_2Ph), 4.92 (d, 2 H, $J = 11.2$ Hz, CH_2Ph), 4.81 (d, 1 H, $J = 1.4$ Hz, H-1), 4.73 (d, 1 H, $J = 11.4$ Hz, CH_2Ph), 4.70–4.63 (m, 5 H, CH_2Ph), 4.61 (d, 1 H, $J = 11.0$ Hz, CH_2Ph), 4.58–4.48 (m, 3 H, CH_2Ph), 4.44 (d, 1 H, $J = 11.5$ Hz, CH_2Ph), 4.17–4.13 (m, 2 H, $\text{CH}_2\text{-CH=CH}_2$), 4.11 (t, 1 H, $J = 9.5$ Hz, H-4''), 4.02 (dd, 1 H, $J = 2.9, 9.5$ Hz, H-3''), 3.99–3.83 (m, 7 H, H-4, H-4', H-3, H-3', H-6'a, H-2', H-6'a''), 3.81 (dd, 1 H, $J = 2.2, 2.4$ Hz, H-2), 3.79–3.65 (m, 8 H, H-6a, PhOCH_3 , H-6'b, H-6b, H-5, H-5'), 3.65–3.51 (m, 3 H, CH_2O , H-5'', H-6'b''), 3.35 (dt, 1 H, $J = 2 \times 6.5, 9.5$ Hz, OCH_2), 3.25 (t, 2 H, $J = 6.9$ Hz, CH_2N_3), 2.15 (s, 3 H, CH_3), 1.72–1.46 (m, 4 H, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$), 1.43–1.22 (m, 8 H, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$), 1.09 (s, 9 H, $\text{C}(\text{CH}_3)_3$); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) $\delta_{\text{C}} = 170.1$ (C=O), 159.3 (Ar), 138.9 (Ar), 138.8 (Ar), 138.6 (Ar), 138.5 (Ar), 138.3 (Ar), 138.0 (Ar), 136.0 (2C, Ar), 135.6 (2C, Ar), 135.3 (Ar), 134.1 ($\text{CH}_2\text{-CH=CH}_2$), 133.3 (Ar), 130.4 (Ar), 129.5 (4 C, Ar), 128.5 (2C, Ar), 128.4 (5 C, Ar), 128.3 (2 C, Ar), 128.2 (2 C, Ar), 128.1 (2 C, Ar), 128.0 (2 C, Ar), 127.7 (5 C, Ar), 127.6 (8 C, Ar), 127.5 (2 C, Ar), 127.3 (3 C, Ar), 127.2 (Ar), 116.9 ($\text{CH}_2\text{-CH=CH}_2$), 113.7 (2 C, Ar), 98.2 (2 C, C-1', C-1''), 97.9 (C-1), 80.5 (C-3), 79.4 (C-3'), 77.9 (C-3''), 75.1 (2 C, CH_2Ph), 74.8 (CH_2Ph), 74.7, 74.6 (C-4, C-4', C-2), 74.3 (C-2'), 74.0 (C-4''), 72.1 (CH_2Ph), 71.7 (2 C, CH_2Ph , C-5''), 71.5 (CH_2Ph), 71.4 ($\text{CH}_2\text{-CH=CH}_2$), 71.2 (2 C, C-5, C-5'), 68.7 (C-2''), 67.6 (CH_2O), 66.2, 66.1 (C-6, C-6'), 62.6 (C-6''), 55.2 (PhOCH_3), 51.5 (CH_2N_3), 29.4, 29.3, 29.1, 28.9 (4 C, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$), 26.8 (3 C, $\text{C}(\text{CH}_3)_3$), 26.7, 26.2 (2 C, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$), 21.2 (CH_3), 19.4 ($\text{C}(\text{CH}_3)_3$). HRMS (ESI) calcd for ($\text{M} + \text{Na}^+$) $\text{C}_{97}\text{H}_{115}\text{N}_3\text{O}_{18}\text{SiNa}$: 1660.7837. Found: 1660.7840.

p-Tolyl 2,3-Di-O-benzoyl- α -D-arabinofuranosyl-(1→5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1→5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1→5)-2,3-di-O-benzoyl-1-thio- α -D-arabinofuranoside (12**).** To a solution of alcohol **18** (258 mg, 0.27 mmol) and trichloroacetimidate (138 μL , 1.37 mmol) in CH_2Cl_2 (3.5 mL) at 0 °C was added DBU (4 μL , 0.027 mmol). The reaction mixture was stirred for 1 h at 0 °C before the solvent was evaporated, and the residue was filtered through a short silica gel column (4:1 hexane–EtOAc, 1% Et_3N). The fractions containing the trichloroacetimidate derivative **19** were evaporated, and the residue was used without any further

purification. The trichloroacetimidate derivative **19** was diluted in CH_2Cl_2 (1 mL) and added to a solution of alcohol **23** (196 mg, 0.17 mmol) in CH_2Cl_2 (1 mL) and 4 Å molecular sieves (86 mg) at -30°C . After stirring for 30 min at -30°C , TMSOTf (3 μL , 0.017 mmol) was added, and the reaction mixture was warmed to 0°C over 30 min, then neutralized by the addition of Et_3N . The solvent was evaporated and the residue was purified by flash chromatography (8.5:1.5 to 7.5:2.5, hexane–EtOAc) to yield **12** (304 mg, 86%) as a white foam. R_f 0.57 (3:2 hexane–EtOAc); $[\alpha]_D + 41.3$ ($c = 0.09$, CH_2Cl_2); $^1\text{H NMR}$ (600 MHz, CDCl_3) $\delta_{\text{H}} = 8.13\text{--}7.85$ (m, 19 H, Ar), 7.74–7.67 (m, 4 H, Ar), 7.63–7.19 (m, 39 H, Ar), 7.10 (d, 2 H, $J = 7.9$ Hz, Ar), 5.76–5.73 (m, 2 H, H-3, H-1), 5.72 (dd, 1 H, $J = 1.5$, 1.8 Hz, H-2), 5.67–5.62 (m, 7 H), 5.57 (br s, 1 H, H-2''), 5.40 (s, 2 H), 5.39 (s, 1 H), 5.38 (s, 1 H), 4.70 (app q, 1 H, $J = 4.0$ Hz, H-4), 4.64–4.58 (m, 3 H, H-4', h-4'', H-4'''), 4.50 (app q, 1 H, $J = 4.1$ Hz, H-4'''), 4.24 (dd, 1 H, $J = 4.2$, 11.2 Hz, H-6a), 4.21–4.15 (m, 3 H), 4.00–3.89 (m, 6 H), 2.31 (s, 3 H, CH_3), 1.02 (s, 9 H, $\text{C}(\text{CH}_3)_3$); $^{13}\text{C NMR}$ (151 MHz, CDCl_3) $\delta_{\text{C}} = 163.9$ (3 \times C=O), 163.8 (2 \times C=O), 163.6 (C=O), 163.5 (2 \times C=O), 163.4 (2 \times C=O), 136.2 (Ar), 134.0 (5 C, Ar), 131.9 (Ar), 131.8 (Ar), 131.7 (2 C, Ar), 131.6 (4 C, Ar), 131.5 (Ar), 131.4 (2 C, Ar), 131.3 (Ar), 130.9 (2 C, Ar), 128.3 (4 C, Ar), 128.2 (10 C, Ar), 128.1 (8 C, Ar), 128.0 (3 C, Ar), 127.7 (Ar), 127.6 (Ar), 127.5 (3 C, Ar), 127.4 (2 C, Ar), 127.3 (Ar), 126.9 (4 C, Ar), 126.8 (6 C, Ar), 126.7 (2 C, Ar), 126.6 (6 C, Ar), 126.5 (2 C, Ar), 126.0 (5 C, Ar), 106.0 (2 C, 2 \times C-1), 105.9 (2 C, 2 \times C-1), 91.6 (C-1), 83.2, 82.1 (4 C), 82.0 (2 C), 81.6 (2 C), 81.5, 77.4 (5 C), 65.9, 65.8 (2 C), 65.7, 63.4, 26.8 (3 C, $\text{C}(\text{CH}_3)_3$), 21.1 (CH_3), 19.3 ($\text{C}(\text{CH}_3)_3$); HRMS (ESI) calcd for ($\text{M} + \text{Na}^+$) $\text{C}_{118}\text{H}_{106}\text{O}_{30}\text{SiNa}$: 2085.6151. Found: 2085.6160.

8-Azidooctyl 2-O-Acetyl-3,4-di-O-benzyl-6-O-(tert-butylidiphenylsilyl)- α -D-mannopyranoside (13). A mixture of trichloroacetimidate **10** (3.89 g, 4.95 mmol), azidoctanol (1.02 g, 5.96 mmol), and 4 Å molecular sieves (2.40 g) in CH_2Cl_2 (45 mL) was stirred for 30 min at -30°C under an argon atmosphere. Then, TMSOTf (135 μL , 0.74 mmol) was added dropwise over 5 min. The reaction mixture was warmed to -5°C over 30 min, and then the TMSOTf was quenched by the addition of Et_3N . The solution was concentrated under vacuum and the resulting syrup was purified by column chromatography (95:5 hexane–EtOAc) to afford **13** (2.97 g, 75%) as a syrup; R_f 0.72 (4:1 hexane–EtOAc); $[\alpha]_D + 14.7$ ($c = 2.76$, CH_2Cl_2); $^1\text{H NMR}$ (500 MHz, CDCl_3) $\delta_{\text{H}} = 7.77$ (d, 2 H, $J = 6.8$ Hz, Ar), 7.73 (d, 2 H, $J = 6.8$ Hz, Ar), 7.49–7.24 (m, 14H, Ar), 7.20 (dd, 2H, $J = 2.83$, 6.7 Hz, Ar), 5.38 (t, 1H, $J = 2.2$ Hz, H-2), 4.93 (d, 1H, $J = 10.7$ Hz, CH_2Ar), 4.83 (d, 1H, $J = 1.7$ Hz, H-1), 4.75 (d, 1H, $J = 11.2$ Hz, CH_2Ar), 4.61 (d, 1H, $J = 10.7$ Hz, CH_2Ar), 4.59 (d, 1H, $J = 11.2$ Hz, CH_2Ar), 4.08–3.97 (m, 3H, H-3, H-4, H-6a), 3.92 (dd, 1H, $J = 11.2$, 1.6 Hz, H-6b), 3.77–3.69 (m, 1H, H-5), 3.66 (dt, 1H, $J = 2 \times 6.8$, 9.6 Hz, OCH_2), 3.40 (dt, 1H, $J = 2 \times 6.4$, 9.6 Hz, OCH_2), 3.25 (t, 2H, $J = 6.7$ Hz, CH_2N_3), 2.16 (s, 3H, OCH_3), 1.66–1.49 (m, 4H, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$), 1.43–1.22 (m, 8H, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$), 1.09 (s, 9H, $\text{C}(\text{CH}_3)_3$); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) $\delta_{\text{C}} = 170.6$ (C=O), 138.5 (Ar), 138.1 (Ar), 136.0 (2C, Ar), 135.6 (2 C, Ar), 133.9 (Ar), 133.3 (Ar), 129.6 (2 C, Ar), 128.4 (2 C, Ar), 128.3 (2 C, Ar), 128.1 (2 C, Ar), 127.9 (2 C, Ar), 127.7 (3 C, Ar), 127.6 (Ar), 127.5 (2 C, Ar), 97.5 (C-1), 78.5 (C-3), 75.4 (CH_2Ph), 74.3 (C-4), 72.7 (C-5), 71.8 (CH_2Ph), 69.2 (C-2), 67.6 (OCH_2), 63.0 (C-6), 51.5 (CH_2N_3), 29.4, 29.3, 29.1, 28.8 (4 C, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$), 26.8 (3 C, $\text{C}(\text{CH}_3)_3$), 26.7, 26.1, (2 C, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$), 21.1 (CH_3), 19.4 ($\text{C}(\text{CH}_3)_3$); HRMS (ESI) calcd for ($\text{M} + \text{Na}^+$) $\text{C}_{46}\text{H}_{59}\text{N}_3\text{O}_7\text{SiNa}$: 816.4014. Found: 816.3999.

8-Azidooctyl 3,4-Di-O-benzyl-2-O-(2-methylnaphthyl)- α -D-mannopyranoside (14). To a solution of **13** (3.32 g, 4.19 mmol) in CH_2Cl_2 – CH_3OH (1:1, 9 mL) was added a solution of NaOCH_3 in CH_3OH (8 mL, 0.1M). The reaction mixture was stirred at room temperature for 1 h, neutralized by the addition of Amberlyst IR-120 (H^+) cation exchange resin, filtered, and concentrated to give a syrup. The crude mixture was dissolved in DMF (20 mL) at 0°C , and sodium hydride (260 mg, 6.52 mmol) and 2-naphthylmethyl bromide (1.06 g, 4.79 mmol) were then added. The mixture was stirred for 3 h at room temperature, diluted with EtOAc, and washed with water (4 \times

20 mL). The organic layers were dried (Na_2SO_4), filtered, and concentrated. The resulting residue was dissolved in THF at 0°C , and the $n\text{-Bu}_4\text{NF}$ (1 M in THF, 20.1 mL) was added. After stirring overnight at room temperature, the reaction mixture was concentrated to give a crude product that was purified by column chromatography (4:1 to 1:1 hexane–EtOAc) to afford **14** (1.36 g, 50% over 3 steps) as a syrup. R_f 0.42 (7:3 hexane–EtOAc); $[\alpha]_D + 16.4$ ($c = 0.23$, CH_2Cl_2); $^1\text{H NMR}$ (500 MHz, CDCl_3) $\delta_{\text{H}} = 7.87\text{--}7.77$ (m, 4 H, Ar), 7.57–7.46 (m, 3 H, Ar), 7.43–7.29 (m, 10 H, Ar), 4.98 (d, 1 H, $J = 10.8$ Hz, CH_2NAP), 4.97 (d, 1 H, $J = 12.6$ Hz, CH_2Ph), 4.88 (d, 1 H, $J = 12.6$ Hz, CH_2Ph), 4.84 (d, 1 H, $J = 1.9$ Hz, H-1), 4.71 (d, 1 H, $J = 11.7$ Hz, CH_2Ph), 4.69 (d, 1 H, $J = 10.8$ Hz, CH_2NAP), 4.68 (d, 1 H, $J = 11.7$ Hz, CH_2Ph), 4.03 (t, 1 H, $J = 9.5$ Hz, H-4), 3.96 (dd, 1 H, $J = 2.9$, 9.5 Hz, H-3), 3.88 (dd, 1 H, $J = 3.1$, 11.7 Hz, H-6a), 3.85 (dd, 1 H, $J = 1.9$, 2.9 Hz, H-2), 3.81 (dd, 1 H, $J = 4.8$, 11.8 Hz, H-6b), 3.67 (ddd, 1 H, $J = 3.0$, 4.6, 9.4 Hz, H-5), 3.62 (dt, 1 H, $J = 2 \times 6.8$, 9.6 Hz, OCH_2), 3.33 (dt, 1 H, $J = 2 \times 6.5$, 9.6 Hz, OCH_2), 3.26 (t, 2 H, $J = 7.0$ Hz, CH_2N_3), 1.74 (br s, 1 H, OH), 1.74 (m, 4 H, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$), 1.43–1.19 (m, 8 H, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) $\delta_{\text{C}} = 138.5$ (Ar), 138.4 (Ar), 135.8 (Ar), 133.2 (Ar), 133.0 (Ar), 128.5 (2 C, Ar), 128.4 (2 C, Ar), 128.2 (Ar), 128.1 (2 C, Ar), 127.9 (Ar), 127.8 (Ar), 127.7 (Ar), 127.6 (3 C, Ar), 126.7 (Ar), 126.1 (Ar), 125.9 (2 C, Ar), 98.3 (C-1), 80.4 (C-3), 75.3 (CH_2NAP), 75.1 (C-2), 74.9 (C-4), 73.0 (CH_2Ph), 72.4 (CH_2Ph), 72.1 (C-5), 67.7 (OCH_2), 62.5 (C-6), 51.5 (CH_2N_3), 29.4, 29.2, 29.0, 28.8, 26.7, 26.0 (6 C, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$). HRMS (ESI) calcd for ($\text{M} + \text{Na}^+$) $\text{C}_{39}\text{H}_{47}\text{N}_3\text{O}_6\text{Na}$: 676.3357. Found: 676.3351.

8-Azidooctyl 2-O-Acetyl-3,4-di-O-benzyl-6-O-(tert-butylidiphenylsilyl)- α -D-mannopyranosyl-(1 \rightarrow 6)-3,4-di-O-benzyl-2-O-(2-methylnaphthyl)- α -D-mannopyranoside (15). A mixture of trichloroacetimidate **10** (118 mg, 0.15 mmol), alcohol **14** (83.3 mg, 0.13 mmol), and 4 Å molecular sieves (50 mg) in CH_2Cl_2 (2 mL) was stirred for 30 min at -20°C under an argon atmosphere. Then, TMSOTf (135 μL , 0.74 mmol) was added dropwise over 5 min. The reaction mixture was warmed to 0°C over 20 min, and then the TMSOTf was quenched by the addition of Et_3N . The solution was concentrated under vacuum and the resulting syrup was purified by column chromatography (95:5 hexane–EtOAc) to afford **15** (134.4 mg, 83%) as a syrup; R_f 0.64 (4:1 hexane–EtOAc); $[\alpha]_D + 28.1$ ($c = 1.90$, CH_2Cl_2); $^1\text{H NMR}$ (500 MHz, CDCl_3) $\delta_{\text{H}} = 7.94\text{--}7.65$ (m, 8 H, Ar), 7.56 (dd, 1 H, $J = 1.2$, 8.3 Hz, Ar), 7.50 7.17 (m, 28 H, Ar), 5.54 (dd, 1 H, $J = 1.9$, 3.1 Hz, H-2'), 5.00 (d, 1 H, $J = 1.9$ Hz, H-1'), 4.98–4.90 (m, 4 H, CH_2NAP , 3 \times CH_2Ph), 4.88 (d, 1 H, $J = 1.6$ Hz, H-1), 4.70–4.61 (m, 4 H, CH_2Ph), 4.53 (d, 1 H, $J = 11.2$ Hz, CH_2NAP), 4.48 (d, 1 H, $J = 11.2$ Hz, CH_2Ph), 4.13 (app t, 1 H, $J = 9.6$ Hz, H-4'), 4.04 (dd, 1 H, $J = 3.1$, 9.6 Hz, H-3'), 4.00–3.81 (m, 6 H, H-3, H-4, H-2, H-6'a, H-6'b, H-6a), 3.76–3.67 (m, 3 H, H-5', H-5, H-6b), 3.62 (dt, 1H, $J = 2 \times 6.8$, 9.5 Hz, OCH_2), 3.35 (dt, 1H, $J = 2 \times 6.5$, 9.5 Hz, OCH_2), 3.23 (t, 2 H, $J = 7.0$ Hz, CH_2N_3), 2.16 (s, 3 H, CH_3), 1.62–1.46 (m, 4 H, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$), 1.40–1.23 (m, 8 H, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$), 1.10 (s, 9 H, $\text{C}(\text{CH}_3)_3$); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) $\delta_{\text{C}} = 170.2$ (C=O), 138.8 (Ar), 138.6 (Ar), 138.4 (Ar), 138.0 (Ar), 136.0 (2 C, Ar), 135.9 (Ar), 135.6 (2 C, Ar), 134.0 (Ar), 133.3 (Ar), 133.2 (Ar), 133.0 (Ar), 129.5 (2 C, Ar), 128.4 (4 C, Ar), 128.3 (6 C, Ar), 128.2 (Ar), 127.9 (Ar), 127.7 (10 C, Ar), 127.6 (Ar), 127.5 (3 C, Ar), 127.4 (Ar), 126.6 (Ar), 126.1 (Ar), 126.0 (Ar), 125.8 (Ar), 98.0 (C-1), 97.6 (C-1'), 80.5 (C-3), 77.9 (C-3'), 75.2 (CH_2NAP), 75.0 (CH_2Ph), 74.8 (C-2), 74.7 (C-4), 74.0 (C-4'), 72.7 (CH_2Ph), 72.6 (C-5), 72.1 (CH_2Ph), 71.5 (CH_2Ph), 71.3 (C-5'), 68.8 (C-2'), 67.5 (OCH_2), 66.5 (C-6), 62.6 (C-6'), 51.4 (CH_2N_3), 29.4, 29.3, 29.1, 28.8 (4 C, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$), 26.8 (3 C, $\text{C}(\text{CH}_3)_3$), 26.6, 26.1 (2 C, $\text{OCH}_2(\text{CH}_2)_6\text{CH}_2\text{N}_3$), 21.1 (CH_3), 19.4 ($\text{C}(\text{CH}_3)_3$). HRMS (ESI) calcd for ($\text{M} + \text{Na}^+$) $\text{C}_{77}\text{H}_{89}\text{N}_3\text{O}_{12}\text{SiNa}$: 1298.6108. Found: 1298.6093.

8-Azidooctyl 2-O-Allyl-3,4-di-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6)-3,4-di-O-benzyl-2-O-(2-methylnaphthyl)- α -D-mannopyranoside (16). To a solution of **15** (1.44 g, 1.13 mmol) in CH_2Cl_2 – CH_3OH (1:1, 6 mL) was added a solution of NaOCH_3 in CH_3OH (2.5 mL, 0.1M). The reaction mixture was stirred for 2 h, neutralized by the addition of Amberlyst IR-120 (H^+) cation exchange

resin, filtered, and concentrated to give a syrup. The crude mixture was dissolved in DMF (6.5 mL) at 0 °C, and sodium hydride (57 mg, 1.41 mmol) and allyl bromide (200 μ L, 4.79 mmol) were then added. The mixture was stirred for 3 h at room temperature, concentrated, diluted with EtOAc, and washed with water (4 \times 20 mL). The organic layers were dried (Na₂SO₄), filtered, and concentrated. The resulting residue was dissolved in THF at 0 °C, and the *n*-Bu₄NF (1 M in THF, 9 mL) was then added. After stirring for 24 h at room temperature, the reaction mixture was concentrated to give a crude product that was purified by column chromatography (9:1 to 7:3 hexane–EtOAc) to afford **16** (1.103 g, 94% over 3 steps) as a syrup. *R*_f 0.24 (8:2 hexane–EtOAc); [α]_D + 30.7 (*c* = 0.60, CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃) δ _H = 7.83–7.72 (m, 4 H, Ar), 7.54 (dd, 1 H, *J* = 1.3, 8.1 Hz, Ar), 7.50–7.44 (m, 2 H, Ar), 7.41–7.18 (m, 20 H, Ar), 5.89 (dddd, 1 H, *J* = 5.0, 5.9, 10.5, 17.2 Hz, CH₂–CH=CH₂), 5.27 (dd, 1 H, *J* = 1.4, 17.2 Hz, CH₂–CH=CH₂), 5.15 (dd, 1 H, *J* = 1.4, 10.5 Hz, CH₂–CH=CH₂), 5.07 (d, 1 H, *J* = 1.3 Hz, H-1'), 4.97–4.86 (m, 4 H, CH₂NAP, 3 \times CH₂Ph), 4.86 (d, 1 H, *J* = 1.3 Hz, H-1), 4.68 (s, 2 H, CH₂Ph), 4.65–4.49 (m, 4 H, CH₂NAP, 3 \times CH₂Ph), 4.18–4.06 (m, 2 H, CH₂–CH=CH₂), 4.02–3.88 (m, 5 H, H-4, H-4', H-3, H-3', H-6'a), 3.86 (m, 2 H, H-2, H-2'), 3.81–3.64 (m, 5 H, H-6a, H-6'b, H-6b, H-5', H-5), 3.60 (dt, 1H, *J* = 2 \times 6.6, 9.6 Hz, OCH₂), 3.34 (dt, 1H, *J* = 2 \times 6.4, 9.6 Hz, OCH₂), 3.24 (t, 2 H, *J* = 7.0 Hz, CH₂N₃), 1.94 (t, 1 H, *J* = 6.0 Hz, OH), 1.64–1.43 (m, 4 H, OCH₂(CH₂)₆CH₂N₃), 1.40–1.27 (m, 8 H, OCH₂(CH₂)₆CH₂N₃); ¹³C NMR (126 MHz, CDCl₃) δ _C = 138.6 (Ar), 138.5 (Ar), 138.5 (Ar), 138.3 (Ar), 135.8 (Ar), 135.0 (CH₂–CH=CH₂), 133.2 (Ar), 133.0 (Ar), 128.4 (6C, Ar), 128.3 (2 C, Ar), 128.2 (Ar), 127.9 (5 C, Ar), 127.8 (2 C, Ar), 127.7 (4 C, Ar), 127.6 (3 C, Ar), 126.7 (Ar), 126.2 (Ar), 126.0 (Ar), 125.9 (Ar), 117.1 (CH₂–CH=CH₂), 98.5 (C-1'), 97.9 (C-1), 80.5 (C-3'), 79.3 (C-3), 75.2 (CH₂NAP), 75.1 (CH₂Ph), 75.0, 74.9 (C-2', C-2), 74.8, 74.7 (C-4', C-4), 73.1 (CH₂Ph), 72.3 (CH₂Ph), 72.2 (C-5'), 72.1 (CH₂–CH=CH₂), 71.7 (CH₂Ph), 71.6 (C-5), 67.6 (OCH₂), 66.1 (C-6'), 62.4 (C-6), 51.4 (CH₂N₃), 29.4, 29.3, 29.1, 28.8, 26.7, 26.1 (6 C, OCH₂(CH₂)₆CH₂N₃); HRMS (ESI) calcd for (M + Na⁺) C₆₂H₇₃N₃O₁₁Na: 1058.5137. Found: 1058.5119.

2,3-Di-O-benzoyl-5-O-(tert-butylidiphenylsilyl)- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranose (18). To a solution of thioglycoside **17** (4.63 g, 6.59 mmol) in THF–H₂O (40:1, 55.5 mL) at 0 °C were added NIS (2.72 g, 12.09 mmol) and AgOTf (673 mg, 2.62 mmol). The reaction mixture was stirred at 0 °C for 3.5 h and then neutralized by the addition of Et₃N. The solvent was evaporated, and the residue was diluted with EtOAc (70 mL) and washed with a satd aq solution of Na₂S₂O₃ (2 \times 50 mL) and water (1 \times 50 mL). The aqueous layers were extracted with EtOAc (2 \times 30 mL), and the combined organic layers were dried (Na₂SO₄) and concentrated. The residue was purified by flash chromatography (4:1 \rightarrow 7.5:2.5, hexane–EtOAc) to yield **18** (3.74 g, 95%, α/β : 4/1) as a white foam. *R*_f 0.56 (7:3 hexane–EtOAc); ¹H NMR (700 MHz, CDCl₃) δ _H = 8.14–7.94 (m, 9 H, Ar), 7.78–7.62 (m, 5 H, Ar), 7.65–7.25 (m, 23 H, Ar), 5.76–5.73 (m, 0.4 H, H-3, H-3'), 5.67–5.62 (m, 3 H, H-1, H-3, H-3'), 5.57 (d, 1 H, *J* = 0.9 Hz, H-2), 5.54 (d, 1 H, *J* = 0.9 Hz, H-2'), 5.52 (d, 0.2 H, *J* = 1.0 Hz, H-2), 5.50 (d, 0.2 H, *J* = 1.0 Hz, H-2'), 5.42 (br s, 0.2 H, H-1), 5.38 (br s, 1 H, H-1'), 5.33 (br s, 0.2 H, H-1'), 4.68 (app q, 1 H, *J* = 4.0 Hz, H-4), 4.60 (app q, 0.2 H, *J* = 4.7 Hz, H-4), 4.52 (app q, 1 H, *J* = 4.7 Hz, H-4'), 4.45 (app q, 0.2 H, *J* = 4.0 Hz, H-4'), 4.23 (dd, 0.2 H, *J* = 2.8, 11.2 Hz, H-6a), 4.19 (dd, 1 H, *J* = 5.0, 11.2 Hz, H-6a), 4.05–3.91 (m, 3.6 H, H-6'a; H-6b, H-6'b, H-6'a; H-6b, H-6'b), 3.03 (d, 1 H, *J* = 3.5 Hz, OH), 1.06 (s, 2 H, C(CH₃)₃), 1.04 (m, 9 H, C(CH₃)₃); ¹³C NMR (126 MHz, CDCl₃) δ _C = 165.9 (2 \times C=O), 165.8 (C=O), 165.7 (C=O), 165.6 (C=O), 165.5 (2 \times C=O), 165.3 (C=O), 135.7 (2 C, Ar), 135.7 (Ar), 133.5 (3 C, Ar), 133.3 (4 C, Ar), 133.2 (4 C, Ar), 133.1 (Ar), 130.1 (Ar), 130.0 (3 C, Ar), 129.9 (4 C, Ar), 129.8 (2 C, Ar), 129.7 (2 C), 129.6, 129.3 (2 C, Ar), 129.2 (2 C, Ar), 129.1 (2 C, Ar), 129.0 (2 C, Ar), 128.5 (2 C, Ar), 128.4 (3 C, Ar), 128.3 (2 C, Ar), 128.1 (Ar), 127.7 (2 C, Ar), 106.6, 106.4, 106.0 (C-1'), 101.0 (C-1), 95.4, 84.6, 83.2 (C-4'), 83.0 (C-4'), 82.8 (C-4), 82.5, 82.4 (C-4), 82.2 (C-2, C-2'), 78.2 (C-3, C-3'), 77.6 (C-3, C-3'), 66.4 (C-6), 66.2 (C-6), 63.4 (C-6'), 63.3 (C-

6'), 26.8 (6 C, 2 \times C(CH₃)₃), 19.3 (2 \times C(CH₃)₃). HRMS (ESI) calcd for (M + Na⁺) C₅₄H₅₂O₁₃SiNa: 959.3069. Found: 959.3073.

***p*-Tolyl 2,3-Di-O-benzoyl-5-O-(tert-butylidiphenylsilyl)- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl-1-thio- α -D-arabinofuranoside (22).** The synthesis of **22** was achieved following the procedure described for the compound **8**, using the disaccharide **20** (3.85 g, 4.79 mmol) and trichloroacetimidate **21** (4 g, 5.39 mmol) in the presence of TMSOTf (78 μ L, 0.43 mmol) in CH₂Cl₂ (60 mL). The product was purified by column chromatography (9:1 \rightarrow 7.5:2.5 hexane–EtOAc) to yield **22** (5.69 g, 86%) as a white foam. *R*_f 0.69 (7:3 hexane–EtOAc); [α]_D + 29.8 (*c* = 1.20, CH₂Cl₂); ¹H NMR (700 MHz, CDCl₃) δ _H = 8.11–7.91 (m, 12 H, Ar), 7.71 (t, 4 H, *J* = 6.9 Hz, Ar), 7.62–7.23 (m, 26 H, Ar), 7.09 (d, 2 H, *J* = 7.9 Hz, Ar), 5.76–5.73 (m, 2 H, H-3, H-1), 5.72 (br s, 1 H, H-2), 5.67–5.62 (m, 3 H, H-3', H-2', H-3''), 5.58 (app s, 1 H, H-2''), 5.39 (app s, 2 H, H-1', H-1''), 4.70 (app q, 1 H, *J* = 4.0 Hz, H-4), 4.63 (app q, 1 H, *J* = 4.2 Hz, H-4'), 4.51 (app q, 1 H, *J* = 4.1 Hz, H-4''), 4.25 (dd, 1 H, *J* = 4.2, 11.3 Hz, H-5a), 4.18 (dd, 1 H, *J* = 4.2, 11.3 Hz, H-5'a), 4.02–3.91 (m, 4 H, H-5b, H-5'b, H-5'a, H-5'b), 2.30 (s, 3 H, CH₃), 1.03 (s, 9 H, C(CH₃)₃); ¹³C NMR (126 MHz, CDCl₃) δ _C = 165.6 (C=O), 165.5 (2 C, 2 \times C=O), 165.3 (C=O), 165.2 (C=O), 165.2 (C=O), 137.9 (Ar), 135.7 (4 C, Ar), 133.5 (2 C, Ar), 133.3 (3 C, Ar), 133.2 (2 C, Ar), 133.0 (Ar), 132.6 (2 C, Ar), 130.0 (4 C, Ar), 129.9 (2 C, Ar), 129.8 (8 C, Ar), 129.6 (2 C, Ar), 129.3 (2 C, Ar), 129.2 (2 C, Ar), 129.1 (Ar), 129.0 (Ar), 128.5 (6 C, Ar), 128.4 (2 C, Ar), 128.3 (2 C, Ar), 128.2 (2 C, Ar), 127.7 (5 C, Ar), 106.0 (2 C, C-1', C-1''), 91.6 (C-1), 83.2 (C-4''), 82.2 (2 C, C-2'', C-4''), 82.1 (C-2), 82.0 (C-4), 81.6 (C-2'), 77.5 (C-3), 77.4 (2 C, C-3', C-3''), 65.8 (2 C, C-6, C-6'), 63.4 (C-6''), 26.7 (3 C, C(CH₃)₃), 21.2 (CH₃), 19.3 (C(CH₃)₃). HRMS (ESI) calcd for (M + Na⁺) C₈₀H₇₄O₁₈SSiNa: 1405.4257. Found: 1405.4265.

***p*-Tolyl 2,3-Di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl-1-thio- α -D-arabinofuranoside (23).** To a solution of trisaccharide **22** (1.27 g, 0.92 mmol) in THF–pyridine (4:1, 8 mL) at 0 °C was added a solution of 70% HF–pyridine (350 μ L) dropwise. The reaction mixture was warmed to room temperature and stirred overnight. After dilution with EtOAc (20 mL), the reaction mixture was poured into a satd aq solution of NaHCO₃ (80 mL) and extracted with EtOAc (2 \times 60 mL). The organic layer was washed with H₂O (2 \times 50), dried (Na₂SO₄), filtered, and concentrated under vacuum to give a syrup that was purified by column chromatography (9:1 \rightarrow 7:3 hexane–EtOAc) to yield **23** (977.5 mg, 93%) as a white foam. *R*_f 0.24 (4:1 hexane–EtOAc); [α]_D + 40.8 (*c* = 0.90, CH₂Cl₂); ¹H NMR (700 MHz, CDCl₃) δ _H = 8.14–8.03 (m, 7 H, Ar), 7.95 (dd, 4 H, *J* = 5.6, 6.9 Hz, Ar), 7.64–7.40 (m, 15 H, Ar), 7.29–7.16 (m, 6 H, Ar), 7.12 (d, 2 H, *J* = 7.9 Hz, Ar), 5.80–5.77 (m, 2 H, H-3, H-1), 5.76 (br s, 1 H, H-2), 5.70–5.66 (m, 3 H, H-3', H-2', H-2''), 5.47 (app d, 1 H, *J* = 4.6 Hz, H-3''), 5.46 (br s, 1 H, H-1'), 5.44 (br s, 1 H, H-1''), 4.73 (app q, 1 H, *J* = 4.0 Hz, H-4), 4.66 (app q, 1 H, *J* = 4.2 Hz, H-4'), 4.52 (app q, 1 H, *J* = 4.1 Hz, H-4''), 4.28 (dd, 1 H, *J* = 4.1, 11.3 Hz, H-5a), 4.21 (dd, 1 H, *J* = 4.4, 11.3 Hz, H-5'a), 4.08–3.95 (m, 4 H, H-5b, H-5'b, H-5'a, H-5'b), 2.45 (dd, 1 H, *J* = 4.8, 8.0 Hz, OH), 2.32 (s, 3 H, CH₃); ¹³C NMR (126 MHz, CDCl₃) δ _C = 166.1 (C=O), 165.7 (C=O), 165.6 (C=O), 165.3 (C=O), 165.2 (C=O), 165.1 (C=O), 138.0 (Ar), 133.6 (2 C, Ar), 133.5 (2 C, Ar), 133.3 (2 C, Ar), 132.6 (2 C, Ar), 130.0 (2 C, Ar), 129.9 (10 C, Ar), 129.8 (2 C, Ar), 129.2 (2 C, Ar), 129.1 (2 C, Ar), 129.0 (Ar), 128.6 (8 C, Ar), 128.4 (4 C, Ar), 128.3 (Ar), 125.4 (Ar), 105.9 (2 C, C-1', C-1''), 91.6 (C-1), 83.8 (C-4''), 82.1 (3 C, C-2, C-4, C-4'), 81.8, 81.7 (C-2'', C-2''), 77.8 (C-3''), 77.5 (C-3), 77.4 (C-3'), 66.2 (C-5'), 65.8 (C-5), 62.4 (C-5''), 21.2 (CH₃). HRMS (ESI) calcd for (M + Na⁺) C₆₄H₅₆O₁₈SiNa: 1167.3080. Found: 1167.3089.

8-Azidoctyl 2-O-Acetyl-3,4-di-O-benzyl-6-O-(tert-butylidiphenylsilyl)- α -D-mannopyranosyl-(1 \rightarrow 6)-2-O-allyl-3,4-di-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6)-3,4-di-O-benzyl-2-O-(2-methylnaphthyl)- α -D-mannopyranoside (24). To a solution of **8** (96 mg, 0.058 mmol) in THF (300 μ L), degassed under vacuum, and stirring under an Ar atmosphere, (1,5-cyclooctadiene)bis-(methylidiphenylphosphine)iridium I hexafluorophosphate catalyst (3 mg, 0.0035 mmol) was added, followed by further degassing of the

osyl-(1→5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1→5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1→2)-3,4-di-O-benzyl-6-O-(*tert*-butyldiphenylsilyl)- α -D-mannopyranosyl-(1→6)-2-O-allyl-3,4-di-O-benzyl- α -D-mannopyranosyl-(1→6)-3,4-di-O-benzyl-2-O-(2-methylnaphthyl)- α -D-mannopyranoside (28). To a solution of **8** (105 mg, 0.063 mmol) in Et₂O–CH₃OH (1:1, 300 μ L) was added a solution of NaOCH₃ in CH₃OH (100 μ L, 0.1M). The reaction mixture was stirred for 3 h, neutralized by the addition of Amberlyst IR-120 (H⁺) cation exchange resin, filtered, and concentrated to give **27** as a syrup. The residue was dissolved in CH₂Cl₂ (2 mL), thioglycoside **12** (125 mg, 0.061 mmol) and 4 Å molecular sieves (19 mg) were added, and the mixture was stirred for 30 min at 0 °C before NIS (19 mg, 0.084 mmol) and AgOTf (4 mg, 0.016 mmol) were added, under an argon atmosphere. The reaction mixture was stirred at 0 °C for 3 h and then neutralized by the addition of Et₃N. The solvent was evaporated, and the residue was diluted with CH₂Cl₂ (10 mL) and washed with a satd aq solution of Na₂S₂O₃ (2 × 10 mL) and water (1 × 10 mL). The aqueous layers were extracted with EtOAc (2 × 15 mL), and the combined organic layers were dried (Na₂SO₄) and concentrated. The residue was purified by flash chromatography (9:1 → 7.5:2.5, hexane–acetone) to yield **28** (156 mg, 73%) as a syrup. *R*_f 0.54 (7:3 hexane–acetone); [α]_D + 30.4 (*c* = 0.46, CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ _H = 8.06–7.82 (m, 21 H, Ar), 7.80–7.62 (m, 12 H, Ar), 7.59–7.03 (m, 74 H, Ar), 5.86 (dddd, 1 H, *J* = 5.4, 6.2, 10.5, 17.2 Hz, CH₂–CH=CH₂), 5.73 (d, 1 H, *J* = 4.8 Hz), 5.71–5.61 (m, 10 H), 5.57 (d, 1 H, *J* = 1.2 Hz), 5.42–5.36 (m, 4 H, 4 × H-1), 5.25 (dd, 1 H, *J* = 1.5, 17.2 Hz, CH₂–CH=CH₂), 5.07 (dd, 1 H, *J* = 1.5, 10.5 Hz, CH₂–CH=CH₂), 5.02 (d, 1 H, *J* = 1.2 Hz, H-1), 5.01 (d, 1 H, *J* = 1.4 Hz, H-1), 4.93–4.83 (m, 6 H, H-1, 5 × CH₂Ph), 4.72 (d, 1 H, *J* = 11.6 Hz, CH₂Ph), 4.67–4.43 (m, 12 H), 4.41 (d, 1 H, *J* = 11.2 Hz), 4.35 (br s, 1 H), 4.23–4.13 (m, 4 H), 4.13–4.04 (m, 3 H), 4.02–3.80 (m, 15 H), 3.73–3.55 (m, 7 H), 3.51–3.46 (m, 1 H), 3.30 (dt, 1 H, *J* = 2 × 6.5, 9.5 Hz, OCH₂), 3.20 (t, 2 H, *J* = 6.9 Hz, CH₂N₃), 1.58–1.44 (m, 4 H, OCH₂(CH₂)₆CH₂N₃), 1.37–1.20 (m, 8 H, OCH₂(CH₂)₆CH₂N₃), 1.02 (br s, 18 H, 2 × C(CH₃)₃); ¹³C NMR (126 MHz, CDCl₃) δ _C = 165.6 (4 C, C=O), 165.5 (C=O), 165.2 (C=O), 165.1 (2 C, C=O), 165.0 (2 C, C=O), 138.9 (Ar), 138.8 (Ar), 138.5 (2 C, Ar), 138.3 (Ar), 138.2 (Ar), 135.9 (2 C, Ar), 135.8 (Ar), 135.7 (3 C, Ar), 135.6 (3 C, Ar), 135.2 (2 C, Ar), 134.1 (Ar), 133.5 (Ar), 133.4 (CH₂–CH=CH₂), 133.3 (2 C, Ar), 133.2 (Ar), 133.1 (5 C, Ar), 130.0 (3 C, Ar), 129.9 (6 C, Ar), 129.8 (12 C, Ar), 129.6 (3 C, Ar), 129.4 (2 C, Ar), 129.3 (2 C, Ar), 129.2 (2 C, Ar), 129.1 (3 C, Ar), 128.5 (4 C, Ar), 128.4 (6 C, Ar), 128.3 (12 C, Ar), 128.2 (10 C, Ar), 128.1 (4 C, Ar), 128.0 (2 C, Ar), 127.9 (2 C, Ar), 127.8 (Ar), 127.7 (11 C, Ar), 127.6 (6 C, Ar), 127.5 (4 C, Ar), 127.3 (Ar), 127.2 (3 C, Ar), 126.7 (Ar), 126.1 (Ar), 126.0 (Ar), 125.9 (Ar), 116.9 (CH₂–CH=CH₂), 106.3 (C-1), 106.0 (3 × C-1), 105.9 (C-1), 99.6, 98.4, 97.9 (C-1, C-1', C-1''), 83.2 (3 C), 82.2, 82.1 (3 C), 81.9, 81.5 (3C), 80.6, 79.9, 79.6, 75.1, 75.0, 74.9, 74.7, 74.6 (2 C), 74.4 (2 C), 74.2, 73.1, 73.0, 72.4, 72.3, 71.9, 71.7, 71.6, 71.4, 71.3, 67.5 (2 C), 66.1, 65.9 (2 C), 65.8 (2 C), 65.7 (2 C), 65.5, 63.4 (2 C), 63.1, 51.4 (CH₂N₃), 29.6, 29.3, 29.0, 28.8 (4 C, OCH₂(CH₂)₆CH₂N₃), 26.9 (6 C, 2 × C(CH₃)₃), 26.7, 26.1 (2 C, OCH₂(CH₂)₆CH₂N₃), 19.4 (C(CH₃)₃), 19.2 (C(CH₃)₃). HRMS (ESI) calcd for (M + 2 Na⁺) C₂₀₉H₂₀₉N₃O₄₆Si₂Na₂: 1800.1746. Found: 1800.1785.

8-Azidooctyl 2,3-Di-O-benzoyl-6-O-(*tert*-butyldiphenylsilyl)- α -D-arabinofuranosyl-(1→5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1→5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1→5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1→2)-3,4-di-O-benzyl-6-O-(*tert*-butyldiphenylsilyl)- α -D-mannopyranosyl-(1→6)-[2,3,4,6-tetra-O-acetyl- α -D-mannopyranosyl-(1→2)]-3,4-di-O-benzyl- α -D-mannopyranosyl-(1→6)-2-O-(2-methylnaphthyl)-3,4-di-O-benzyl- α -D-mannopyranoside (29). The synthesis of **29** was achieved following the procedure described for the preparation of **24**, starting from the octasaccharide **28** (93 mg, 0.026 mmol) and using (1,5-cyclooctadiene)bis(methyldiphenylphosphine) iridium I hexafluorophosphate catalyst (3 mg, 0.0035 mmol) in THF (500 μ L), then HgO (8 mg, 0.037 mmol) and HgCl₂ (9 mg, 0.033 mmol) in acetone–water (10:1, 1.8 mL). The crude residue, used without any further purification, was dissolved in CH₂Cl₂ (600 μ L). Then, **11a** (36

mg, 0.079 mmol) and 4 Å molecular sieves (18 mg) were added and the mixture was stirred for 30 min at 0 °C. NIS (19 mg, 0.084 mmol) and AgOTf (4 mg, 0.016 mmol) were then added, under an argon atmosphere. The reaction mixture was stirred at 0 °C for 2 h, then neutralized by the addition of Et₃N. The solvent was evaporated, and the residue was diluted with CH₂Cl₂ (10 mL) and washed with a satd aq solution of Na₂S₂O₃ (2 × 10 mL) and water (1 × 10 mL). The aqueous layers were extracted with EtOAc (2 × 15 mL), and the combined organic layers were dried (Na₂SO₄) and concentrated. The residue was purified by gel filtration chromatography (Sephadex, LH-20) with 1:1 CH₂Cl₂–CH₃OH as the eluent, to yield **29** (88.5 mg, 88% over 2 steps). *R*_f 0.34 (4:1, hexane–EtOAc); [α]_D + 1.3 (*c* = 0.03, CH₂Cl₂); ¹H NMR (500 MHz, CDCl₃) δ _H = 8.05–7.81 (m, 20 H, Ar), 7.80–7.62 (m, 12 H, Ar), 7.58–7.01 (m, 75 H, Ar), 5.76 (d, 1 H, *J* = 4.9 Hz), 5.72 (d, 1 H, *J* = 1.2 Hz), 5.71–5.60 (m, 8 H), 5.56 (br s, 1 H), 5.46 (dd, 1 H, *J* = 1.6, 3.2 Hz), 5.43–5.35 (m, 5 H), 5.28 (t, 1 H, *J* = 10.1 Hz), 5.01 (s, 3 H), 4.95–4.83 (m, 6 H), 4.78 (d, 1 H, *J* = 11.7 Hz, CH₂Ph), 4.67–4.45 (m, 13 H), 4.37 (br s, 1 H), 4.28 (dd, 1 H, *J* = 4.7, 11.9 Hz), 4.22–4.08 (m, 8 H), 4.03–3.79 (m, 14 H), 3.74 (m, 3 H), 3.68 (dd, 1 H, *J* = 2.9, 9.7 Hz), 3.64–3.52 (m, 3 H), 3.45 (d, 1 H, *J* = 10.4 Hz), 3.30 (dt, 1 H, *J* = 2 × 6.5, 9.5 Hz, OCH₂), 3.20 (t, 2 H, *J* = 6.9 Hz, CH₂N₃), 2.02 (s, 6 H, 2 × CH₃), 1.88 (s, 3 H, CH₃), 1.81 (s, 3 H, CH₃), 1.58–1.43 (m, 4 H, OCH₂(CH₂)₆CH₂N₃), 1.35–1.18 (m, 8 H, OCH₂(CH₂)₆CH₂N₃), 1.02 (s, 9 H, C(CH₃)₃), 1.01 (s, 9 H, C(CH₃)₃); ¹³C NMR (126 MHz, CDCl₃) δ _C = 170.6 (C=O), 170.0 (C=O), 169.6 (C=O), 169.4 (C=O), 165.6 (4 C, C=O), 165.5 (C=O), 165.2 (C=O), 165.1 (2 C, C=O), 165.0 (C=O), 164.9 (C=O), 139.0 (Ar), 138.7 (Ar), 138.5 (2 C, Ar), 138.4 (Ar), 137.9 (Ar), 135.9 (2 C, Ar), 135.8 (Ar), 135.7 (3 C, Ar), 135.6 (3 C, Ar), 134.0 (Ar), 133.5 (Ar), 133.3 (2 C, Ar), 133.2 (Ar), 133.1 (3 C, Ar), 133.0 (2 C, Ar), 130.0 (3 C, Ar), 129.9 (6 C, Ar), 129.8 (12 C, Ar), 129.7 (3 C, Ar), 129.4 (2 C, Ar), 129.3 (2 C, Ar), 129.2 (2 C, Ar), 129.1 (3 C, Ar), 128.5 (4 C, Ar), 128.4 (6 C, Ar), 128.3 (12 C, Ar), 128.2 (10 C, Ar), 128.1 (4 C, Ar), 128.0 (2 C, Ar), 127.9 (2 C, Ar), 127.8 (Ar), 127.7 (12 C, Ar), 127.6 (6 C, Ar), 127.5 (5 C, Ar), 127.2 (3 C, Ar), 127.1 (Ar), 126.6 (Ar), 126.1 (Ar), 125.9 (Ar), 125.9 (Ar), 106.5 (C-1), 106.0 (3 × C-1), 105.9 (C-1), 99.6 (2 × C-1), 99.2 (C-1), 97.9 (C-1), 83.2 (3 C), 82.2, 82.0 (3 C), 81.9, 81.6, 81.5 (3 C), 81.4, 80.6, 80.1, 79.4, 78.5, 76.0, 75.1, 75.0, 74.9, 74.5 (2 C), 74.4 (2 C), 74.3, 73.2, 72.8 (2 C), 72.3 (2 C), 71.8 (2 C), 71.6, 71.5, 71.1, 70.9, 69.5, 69.1 (2 C), 68.9, 67.6 (2 C), 67.2, 66.0, 65.9 (2 C), 65.8 (2 C), 65.7 (2 C), 63.5 (2 C), 63.1, 62.4 (2 C), 51.4 (CH₂N₃), 29.7, 29.4, 29.0, 28.8 (4 C, OCH₂(CH₂)₆CH₂N₃), 26.8 (6 C, 2 × C(CH₃)₃), 26.6, 26.1 (2 C, OCH₂(CH₂)₆CH₂N₃), 20.8 (CH₃), 20.7 (CH₃), 20.6 (CH₃), 20.5 (CH₃), 19.3 (C(CH₃)₃), 19.2 (C(CH₃)₃); HRMS (ESI) calcd for (M + 2 Na⁺) C₂₂₀H₂₂₅N₃O₅₅Si₂Na₂: 1945.2112. Found: 1945.2137.

8-Azidooctyl 2-O-Acetyl-3,4-di-O-benzyl-6-O-(*tert*-butyldiphenylsilyl)- α -D-mannopyranosyl-(1→6)-2-O-allyl-3,4-di-O-benzyl- α -D-mannopyranosyl-(1→6)-3,4-di-O-benzyl- α -D-mannopyranoside (30). To a solution of **9** (48 mg, 0.029 mmol) in CH₃CN–H₂O (10:1, 440 μ L) was added CAN (32 mg, 0.058 mmol). After 1 h stirring, the reaction mixture was concentrated under vacuum. The residue was diluted with EtOAc (15 mL) and washed with an aq solution of NaHCO₃ (2 × 10 mL) and water (1 × 10 mL). The aqueous layers were extracted with EtOAc (2 × 10 mL), and the combined organic layers were dried (Na₂SO₄) and concentrated. The residue was purified by flash chromatography (4:1 → 7:3, hexane–EtOAc) to yield **30** (37 mg, 83%) as a syrup. *R*_f 0.71 (3:2 hexane–EtOAc); [α]_D + 40.5 (*c* = 0.50, CH₂Cl₂); ¹H NMR (500 MHz, CDCl₃) δ _H = 7.76 (d, 2 H, *J* = 6.8 Hz, Ar), 7.68 (d, 2 H, *J* = 6.8 Hz, Ar), 7.47–7.10 (m, 36 H, Ar), 5.95 (dddd, 1 H, *J* = 5.4, 6.2, 10.5, 17.2 Hz, CH₂–CH=CH₂), 5.51 (dd, 1 H, *J* = 1.7, 2.9 Hz, H-2''), 5.34 (dd, 1 H, *J* = 1.4, 17.2 Hz, CH₂–CH=CH₂), 5.16 (dd, 1 H, *J* = 1.4, 10.5 Hz, CH₂–CH=CH₂), 5.02 (d, 1 H, *J* = 1.3 Hz, H-1'), 4.96–4.89 (m, 3 H, H-1'', 2 × CH₂Ph), 4.88–4.83 (m, 2 H, H-1, CH₂Ph), 4.76–4.68 (m, 4 H, CH₂Ph), 4.64 (d, 1 H, *J* = 11.8 Hz, CH₂Ph), 4.61 (d, 1 H, *J* = 10.9 Hz, CH₂Ph), 4.66–4.58 (m, 2 H, CH₂Ph), 4.46 (d, 1 H, *J* = 11.4 Hz, CH₂Ph), 4.21–4.16 (m, 2 H, CH₂–CH=CH₂), 4.11 (t, 1 H, *J* = 9.6 Hz, H-4''), 4.05 (br s, 1 H, H-2), 4.02 (dd, 1 H, *J* = 2.9, 9.6 Hz, H-

3''), 3.94–3.85 (m, 5 H, H-4, H-3, H-3', H-6a, H-6''a), 3.83 (br s, 1 H, H-2'), 3.78–3.54 (m, 9 H, H-4', H-6''b, H-6b, H-6'a, H-6'b, H-5, H-5', H-5'', CH₂O), 3.40 (dt, 1 H, *J* = 2 × 6.5, 9.5 Hz, OCH₂), 3.25 (t, 2 H, *J* = 6.9 Hz, CH₂N₃), 2.46 (d, *J* = 2.4 Hz, 1 H), 2.15 (s, 3 H, CH₃), 1.64–1.48 (m, 4 H, OCH₂(CH₂)₆CH₂N₃), 1.41–1.25 (m, 8 H, OCH₂(CH₂)₆CH₂N₃), 1.08 (s, 9 H, C(CH₃)₃); ¹³C NMR (126 MHz, CDCl₃) δ_C = 170.3 (C=O), 138.9 (Ar), 138.7 (Ar), 138.4 (Ar), 138.3 (Ar), 138.0 (Ar), 137.9 (Ar), 136.0 (2C, Ar), 135.5 (2C, Ar), 135.2 (Ar), 134.0 (CH₂–CH=CH₂), 133.3 (Ar), 129.5 (2C, Ar), 128.6 (2C, Ar), 128.4 (7 C, Ar), 128.2 (2 C, Ar), 128.1 (2 C, Ar), 128.0 (Ar), 127.9 (4 C, Ar), 127.7 (5 C, Ar), 127.6 (5 C, Ar), 127.5 (2 C, Ar), 127.3 (3 C, Ar), 127.2 (Ar), 117.2 (CH₂–CH=CH₂), 99.0 (C-1), 98.2 (C-1'), 97.8 (C-1''), 80.5 (C-3), 79.7 (C-3'), 77.9 (C-3''), 75.1 (2 C, CH₂Ph), 74.8 (CH₂Ph), 74.6, 74.4 (C-4, C-4', C-2), 74.1 (C-2''), 74.0 (C-4''), 72.5 (C-5''), 72.0 (CH₂Ph), 71.8 (2 C, CH₂Ph), 71.5 (CH₂–CH=CH₂), 71.1, 70.9 (C-5, C-5'), 68.7 (C-2''), 68.4 (C-2), 67.6 (CH₂O), 66.2 (2 C, C-6, C-6'), 62.6 (C-6''), 51.4 (CH₂N₃), 29.4, 29.3, 29.1, 28.8 (4 C, OCH₂(CH₂)₆CH₂N₃), 26.8 (3 C, C(CH₃)₃), 26.7, 26.1 (2 C, OCH₂(CH₂)₆CH₂N₃), 21.1 (CH₃), 19.4 (C(CH₃)₃); HRMS (ESI) calcd for (M + Na⁺) C₃₉H₁₀₇N₃O₁₇SiNa: 1540.7262. Found: 1540.7256.

8-Azidoethyl 3,4-Di-O-benzyl-2-O-(4-methoxybenzyl)-α-D-mannopyranoside (31). The synthesis of 31 (2.86 g, 3.60 mmol) was achieved following the procedure described for the preparation of 14, using NaOCH₃ in CH₃OH (7 mL, 0.1M), CH₂Cl₂–CH₃OH (1:1, 8 mL), sodium hydride (180 mg, 7.5 mmol), *p*-methoxybenzyl chloride (735 μL, 5.4 mmol), DMF (20 mL), *n*-Bu₄NF (1 M in THF, 28 mL), and THF (9 mL). The product was purified by column chromatography (4:1 to 1:1 hexane–EtOAc) to afford 31 (1.62 g, 93% over 3 steps) as a syrup. *R*_f 0.23 (4:1 hexane–EtOAc); [α]_D²⁰ + 12.4 (*c* = 0.10, CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃) δ_H = 7.41–7.23 (m, 12 H, Ar), 6.85 (d, 2 H, *J* = 8.7 Hz, Ph–OCH₃), 4.94 (d, 1 H, *J* = 10.9 Hz, CH₂Ph), 4.76 (d, 1 H, *J* = 1.0 Hz, H-1), 4.72 (d, 1 H, *J* = 12.0 Hz, CH₂Ph), 4.68–4.59 (m, 4 H, CH₂Ph), 3.95 (app t, 1 H, *J* = 9.2 Hz, H-4), 3.90 (dd, 1 H, *J* = 2.8, 9.2 Hz, H-3), 3.87–3.73 (m, 6 H, H-6a, PhOCH₃, H-6b, H-2), 3.68–3.56 (m, 2 H, H-5, OCH₂), 3.32 (dt, 1 H, *J* = 2 × 6.6, 9.5 Hz, OCH₂), 3.25 (t, 2 H, *J* = 6.9 Hz, OCH₂N₃), 2.01 (t, 1 H, *J* = 6.4 Hz, OH), 1.66–1.43 (m, 4 H, OCH₂(CH₂)₆CH₂N₃), 1.43–1.23 (m, 8 H, OCH₂(CH₂)₆CH₂N₃); ¹³C NMR (126 MHz, CDCl₃) δ_C = 159.3 (Ar), 138.6 (Ar), 138.4 (Ar), 130.4 (Ar), 129.5 (2C, Ar), 128.5 (2 C, Ar), 128.4 (2 C, Ar), 128.1 (2 C, Ar), 127.8 (Ar), 127.6 (2 C, Ar), 127.5 (Ar), 113.7 (2 C, Ar), 98.3 (C-1), 80.3 (C-3), 75.3 (CH₂Ph), 75.1 (C-4), 74.4 (C-2), 72.5 (CH₂Ph), 72.2 (CH₂Ph), 72.0 (C-5), 67.6 (CH₂O), 62.5 (C-6), 55.3 (OCH₃), 51.5 (CH₂N₃), 29.4, 29.3, 29.1, 28.8, 26.7, 26.0 (6 C, OCH₂(CH₂)₆CH₂N₃); HRMS (ESI) calcd for (M + Na⁺) C₃₆H₄₇N₃O₇Na: 656.3306. Found: 656.3303.

8-Azidoethyl 2-O-Acetyl-3,4-di-O-benzyl-6-O-(tert-butylidiphenylsilyl)-α-D-mannopyranosyl-(1→6)-3,4-di-O-benzyl-2-O-(4-methoxybenzyl)-α-D-mannopyranoside (32). The synthesis of 32 was achieved following the procedure described for the preparation of 15, using alcohol 31 (37.6 mg, 0.059 mmol) and trichloroacetimidate 10 (58 mg, 0.074 mmol) in the presence of TMSOTf (130 μL of a 0.07 M solution in Et₂O) in Et₂O (670 μL). The crude residue was purified by column chromatography (9:1 → 8.5:1.5 hexane–EtOAc) to yield 32 (70 mg, 95%) as a syrup. *R*_f 0.61 (8:2 hexane–EtOAc); [α]_D²⁰ + 23.0 (*c* = 0.57, CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃) δ_H = 7.79–7.73 (m, 2 H, Ar), 7.72–7.66 (m, 2 H, Ar), 7.46–7.13 (m, 28 H, Ar), 6.82 (d, 2 H, *J* = 8.5 Hz, PhOCH₃), 5.51 (dd, 1 H, *J* = 1.9, 2.8 Hz, H-2'), 4.96 (d, 1 H, *J* = 1.9 Hz, H-1'), 4.94 (d, 1 H, *J* = 11.5 Hz, CH₂Ph), 4.90 (d, 1 H, *J* = 11.2 Hz, CH₂Ph), 4.80 (d, 1 H, *J* = 1.0 Hz, H-1), 4.69 (d, 1 H, *J* = 11.5 Hz, CH₂Ph), 4.65–4.58 (m, 5 H, CH₂Ph), 4.52–4.45 (m, 2 H, CH₂Ph), 4.10 (t, 1 H, *J* = 9.4 Hz, H-4'), 4.01 (dd, 1 H, *J* = 2.7, 9.4 Hz, H-3'), 3.94 (dd, 1 H, *J* = 2.9, 11.1 Hz, H-6'a), 3.90–3.73 (m, 8 H, H-3, H-4, H-6a, H-6'b, H-2, PhOCH₃), 3.72–3.63 (m, 3 H, H-5, H-5', H-6b), 3.63–3.57 (m, 1 H, OCH₂), 3.34 (dt, 1 H, *J* = 2 × 6.4, 9.5 Hz, OCH₂), 3.23 (t, 2 H, *J* = 6.9 Hz, OCH₂N₃), 2.15 (s, 3 H, CH₃), 1.63–1.45 (m, 4 H, OCH₂(CH₂)₆CH₂N₃), 1.41–1.23 (m, 8 H), 1.07 (s, 9 H, C(CH₃)₃); ¹³C NMR (126 MHz, CDCl₃) δ_C = 170.3 (C=O), 159.3 (Ar), 138.8

(Ar), 138.6 (Ar), 138.5 (Ar), 138.0 (Ar), 136.0 (Ar), 135.5 (3 C, Ar), 134.0 (Ar), 133.3 (Ar), 130.5 (Ar), 129.6 (2 C, Ar), 129.4 (3 C, Ar), 128.5 (2 C, Ar), 128.4 (2 C, Ar), 128.3 (4 C, Ar), 128.2 (2 C, Ar), 127.7 (4 C, Ar), 127.6 (4 C, Ar), 127.5 (3 C, Ar), 127.5 (Ar), 127.4 (Ar), 113.7 (2 C, Ar), 98.0 (C-1'), 97.6 (C-1), 80.4 (C-3), 77.9 (C-3'), 75.2 (CH₂Ph), 75.0 (CH₂Ph), 74.7 (C-4), 74.3 (C-4'), 74.0 (C-2), 72.6 (C-5), 72.1 (CH₂Ph), 71.9 (CH₂Ph), 71.5 (CH₂Ph), 71.2 (C-5'), 68.7 (C-2'), 67.5 (CH₂O), 66.6 (C-6), 62.7 (C-6'), 55.3 (OCH₃), 51.5 (CH₂N₃), 29.4, 29.3, 29.1, 28.9 (4 C, OCH₂(CH₂)₆CH₂N₃), 26.9 (3 C, C(CH₃)₃), 26.7, 26.2 (2 C, OCH₂(CH₂)₆CH₂N₃), 21.1 (CH₃), 19.4 (C(CH₃)₃); HRMS (ESI) calcd for (M + Na⁺) C₇₄H₈₉N₃O₁₃SiNa: 1278.6057. Found: 1278.6055.

8-Azidoethyl 2-O-Allyl-3,4-di-O-benzyl-α-D-mannopyranosyl-(1→6)-3,4-di-O-benzyl-2-O-(4-methoxybenzyl)-α-D-mannopyranoside (33). The synthesis of 33 (2.27 g, 1.8 mmol) was achieved following the procedure described for the preparation of 16 using NaOCH₃ in CH₃OH (4 mL, 0.1M), CH₂Cl₂–CH₃OH (1:1, 6 mL), sodium hydride (92 mg, 3.8 mmol), allyl bromide (310 μL, 3.7 mmol), DMF (10 mL), *n*-Bu₄NF (1 M in THF, 18 mL), and THF (6 mL). The product was purified by column chromatography (9:1 to 3:2 hexane–EtOAc) to afford 33 (1.33 g, 73% over 3 steps) as a syrup. *R*_f 0.32 (3:2 hexane–EtOAc); [α]_D²⁰ + 18.2 (*c* = 0.09, CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ_H = 7.43–7.20 (m, 22 H, Ar), 6.85 (d, 2 H, *J* = 8.6 Hz, PhOCH₃), 5.93 (dddd, 1 H, *J* = 5.0, 5.6, 10.5, 17.2 Hz, CH₂–CH=CH₂), 5.31 (dd, 1 H, *J* = 1.2, 17.2 Hz, CH₂–CH=CH₂), 5.19 (dd, 1 H, *J* = 1.2, 10.6 Hz, CH₂–CH=CH₂), 5.07 (d, 1 H, *J* = 1.1 Hz, H-1'), 4.95 (d, 1 H, *J* = 11.2 Hz, CH₂Ph), 4.93 (d, 1 H, *J* = 11.2 Hz, CH₂Ph), 4.80 (d, 1 H, *J* = 1.1 Hz, H-1), 4.73–4.58 (m, 7 H, CH₂Ph), 4.52 (d, 1 H, *J* = 11.0 Hz, CH₂Ph), 4.20–4.10 (m, 2 H, CH₂–CH=CH₂), 3.97–3.86 (m, 6 H, H-4, H-4', H-3, H-3', H-6a, H-2'), 3.82–3.75 (m, 5 H, H-2, PhOCH₃, H-6'a), 3.74–3.65 (m, 4 H, H-6b, H-6'b, H-5, H-5'), 3.61 (dt, 1 H, *J* = 2 × 6.8, 9.5 Hz, OCH₂), 3.36 (dt, 1 H, *J* = 2 × 6.5, 9.5 Hz, OCH₂), 3.26 (t, 2 H, *J* = 7.0 Hz, CH₂N₃), 1.93 (br s, 1 H, OH), 1.66–1.49 (m, 4 H, OCH₂(CH₂)₆CH₂N₃), 1.41–1.26 (m, 8 H, OCH₂(CH₂)₆CH₂N₃); ¹³C NMR (126 MHz, CDCl₃) δ_C = 159.3 (Ar), 138.6 (Ar), 138.6 (Ar), 138.5 (Ar), 138.3 (Ar), 135.1 (CH₂–CH=CH₂), 130.4 (Ar), 129.5 (2 C, Ar), 128.4 (5 C, Ar), 128.3 (3 C, Ar), 127.9 (4 C, Ar), 127.8 (3 C, Ar), 127.6 (10 C, Ar), 117.1 (CH₂–CH=CH₂), 113.7 (2 C, Ar), 98.4 (C-1'), 97.8 (C-1), 80.4 (C-3), 79.3 (C-3'), 75.1 (2 C, CH₂Ph), 74.9 (C-2'), 74.8, 74.6 (C-4, C-4'), 74.5 (C-2), 72.5 (CH₂Ph), 72.2 (C-5), 72.1 (CH₂Ph), 71.7 (CH₂Ph), 71.6 (C-5'), 67.6 (CH₂O), 66.1 (C-6), 62.4 (C-6'), 55.2 (PhCH₃), 51.4 (CH₂N₃), 29.4, 29.3, 29.1, 28.8, 26.7, 26.1 (6 C, OCH₂(CH₂)₆CH₂N₃). HRMS (ESI) calcd for (M + Na⁺) C₃₉H₇₃N₃O₁₂Na: 1038.5086. Found: 1038.5093.

8-Azidoethyl 2-O-Acetyl-3,4-di-O-benzyl-6-O-(tert-butylidiphenylsilyl)-α-D-mannopyranosyl-(1→6)-2-O-allyl-3,4-di-O-benzyl-α-D-mannopyranosyl-(1→6)-[2,3-di-O-benzoyl-6-O-(tert-butylidiphenylsilyl)-α-D-arabinofuranosyl-(1→5)-2,3-di-O-benzoyl-α-D-arabinofuranosyl-(1→5)-2,3-di-O-benzoyl-α-D-arabinofuranosyl-(1→5)-2,3-di-O-benzoyl-α-D-arabinofuranosyl-(1→2)]-3,4-di-O-benzyl-α-D-mannopyranoside (34). The synthesis of 34 was achieved following the procedure described for the preparation of 25, using alcohol 30 (86 mg, 0.057 mmol), thioglycoside 12 (140 mg, 0.068 mmol), and 4 Å molecular sieves (21 mg) in CH₂Cl₂ (800 μL) in the presence of NIS (22 mg, 0.098 mmol) and AgOTf (4 mg, 0.016 mmol). The product residue was purified by column chromatography (8.5:1.5 → 7.5:2.5, hexane–acetone) to yield 34 (166 mg, 85%) as a syrup. *R*_f 0.43 (7:3 hexane–acetone); [α]_D²⁰ + 27.2 (*c* = 0.15, CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ_H = 8.06–7.85 (m, 20 H, Ar), 7.78–7.64 (m, 8 H, Ar), 7.60–7.06 (m, 73 H, Ar), 5.90 (dddd, 1 H, *J* = 5.4, 6.2, 10.5, 17.2 Hz, CH₂–CH=CH₂), 5.75–5.61 (m, 10 H), 5.58 (br s, 1 H), 5.54 (app t, 1 H, *J* = 2.1 Hz, H-2''), 5.44–5.36 (m, 4 H), 5.28 (dd, 1 H, *J* = 1.5, 17.2 Hz, CH₂–CH=CH₂), 5.10 (dd, 1 H, *J* = 1.5, 10.5 Hz, CH₂–CH=CH₂), 5.05 (d, 1 H, *J* = 1.2 Hz, H-1'), 4.97–4.82 (m, 5 H, H-1'', H-1, 3 × CH₂Ph), 4.76–4.65 (m, 3H), 4.65–4.44 (m, 10 H), 4.42 (d, 1 H, *J* = 11.4 Hz, CH₂Ph), 4.24 (s, 1 H), 4.22–4.08 (m, 7 H), 4.05–3.59 (m, 20 H), 3.56 (d, 1 H, *J* = 9.4 Hz), 3.50 (d, *J* = 10.7 Hz, 1 H), 3.36 (dt, 1 H, *J* = 2 × 6.5, 9.5 Hz, OCH₂), 3.21 (t, 2 H, *J* = 7 Hz, CH₂N₃), 2.15 (s, 3 H, CH₃), 1.61–1.48 (m, 4 H,

OCH₂(CH₂)₆CH₂N₃), 1.39–1.22 (m, 8 H, OCH₂(CH₂)₆CH₂N₃), 1.08 (s, 9 H, C(CH₃)₃), 1.03 (s, 9 H, C(CH₃)₃); ¹³C NMR (126 MHz, CDCl₃) δ_C = 170.1 (C=O), 165.6 (4 C, C=O), 165.5 (C=O), 165.2 (2 C, C=O), 165.1 (3 C, C=O), 138.9 (Ar), 138.8 (Ar), 138.4 (2 C, Ar), 138.2 (Ar), 138.0 (Ar), 136.0 (3 C, Ar), 135.7 (7 C, Ar), 135.6 (3 C, Ar), 135.2 (Ar), 134.0 (Ar), 133.5 (CH₂–CH=CH₂), 133.4 (2 C, Ar), 133.3 (Ar), 133.2 (Ar), 133.1 (2 C, Ar), 133.0 (Ar), 130.0 (3 C, Ar), 129.9 (8 C, Ar), 129.8 (7 C, Ar), 129.6 (2 C, Ar), 129.5 (2 C, Ar), 129.3 (3 C, Ar), 129.2 (3 C, Ar), 129.1 (2 C, Ar), 128.5 (9 C, Ar), 128.4 (9 C, Ar), 128.3 (8 C, Ar), 128.2 (7 C, Ar), 128.1 (2 C, Ar), 127.8 (4 C, Ar), 127.7 (9 C, Ar), 127.6 (3 C, Ar), 127.5 (7 C, Ar), 127.3 (3 C, Ar), 127.2 (Ar), 116.9 (CH₂–CH=CH₂), 106.4, 106.0 (2 C), 105.9 (2 C), 99.4 (C-1), 98.3 (C-1'), 97.9 (C-1'), 83.2 (2 C), 82.1 (6 C), 82.0, 81.6 (2 C), 81.5 (3 C), 80.6, 80.2, 77.7 (2 C), 75.1 (2 C), 74.8, 74.7 (2 C), 74.2, 73.9, 72.5 (2 C), 72.1, 72.0, 71.7, 71.5, 71.4, 71.3, 68.6, 67.6 (CH₂O), 66.3, 66.1, 65.9, 65.8, 65.7, 63.4, 62.6, 60.4, 51.4 (CH₂N₃), 29.5, 29.4, 29.1, 28.8 (4 C, OCH₂(CH₂)₆CH₂N₃), 26.8 (6 C, 2 × C(CH₃)₃), 26.7, 26.2 (2 C, OCH₂(CH₂)₆CH₂N₃), 21.1 (CH₃), 19.4 (C(CH₃)₃), 19.3 (C(CH₃)₃). HRMS (ESI) calcd for (M + 2 Na⁺) C₂₀₀H₂₀₅N₃O₄₇Si₂Na₂: 1751.1533. Found: 1751.1559.

8-Azidooctyl 3,4-Di-O-benzyl-6-O-(tert-butyl)diphenylsilyl)-α-D-mannopyranosyl-(1→6)-2-O-allyl-3,4-di-O-benzyl-α-D-mannopyranosyl-(1→6)-3,4-di-O-benzyl-α-D-mannopyranoside (35). To a solution of **30** (35 mg, 0.023 mmol) in Et₂O–CH₃OH (1:1, 400 μL) was added a solution of NaOCH₃ in CH₃OH (100 μL, 0.1M). The reaction mixture was stirred for 3 h, neutralized by the addition of Amberlyst IR-120 (H⁺) cation exchange resin, filtered, and concentrated to give a syrup that was purified by column chromatography (4:1 hexane–EtOAc) to afford **35** (33 mg, 97%) as a syrup. R_f 0.55 (3:2 hexane–EtOAc); [α]_D + 40.9 (c = 0.16, CH₂Cl₂); ¹H NMR (500 MHz, CDCl₃) δ_H = 7.76 (d, 2 H, J = 6.8 Hz, Ar), 7.69 (d, 2 H, J = 6.8 Hz, Ar), 7.45–7.12 (m, 36 H, Ar), 5.95 (dddd, 1 H, J = 5.4, 6.2, 10.5, 17.2 Hz, CH₂–CH=CH₂), 5.33 (dd, 1 H, J = 1.5, 17.2 Hz, CH₂–CH=CH₂), 5.16 (dd, 1 H, J = 1.5, 10.5 Hz, CH₂–CH=CH₂), 5.05 (d, 1 H, J = 1.2 Hz, H-1''), 4.99 (d, 1 H, J = 1.4 Hz, H-1'), 4.92–4.84 (m, 4 H, H-1, 3 × CH₂Ph), 4.76–4.59 (m, 7 H, CH₂Ph), 4.51 (d, 1 H, J = 11.1 Hz, CH₂Ph), 4.50 (d, 1 H, J = 11.2 Hz, CH₂Ph), 4.20–4.15 (m, 3 H, CH₂–CH=CH₂, H-2''), 4.06 (br s, 1 H, H-2), 3.99 (t, 1 H, J = 9.6 Hz, H-4''), 3.94–3.61 (m, 16 H, H-4, H-3, H-3', H-3'', H-6a, H-6'a, H-2', H-4', H6''b, H-6b, H-6'a, H-6'b, H-5, H-5', H-5'', CH₂O), 3.40 (dt, 1 H, J = 2 × 6.5, 9.5 Hz, OCH₂), 3.24 (t, 2 H, J = 6.9 Hz, CH₂N₃), 2.62 (d, J = 2.6 Hz, 1 H, OH), 2.54 (br s, 1 H, OH), 1.63–1.48 (m, 4 H, OCH₂(CH₂)₆CH₂N₃), 1.40–1.25 (m, 8 H, OCH₂(CH₂)₆CH₂N₃), 1.06 (s, 9 H, C(CH₃)₃); ¹³C NMR (126 MHz, CDCl₃) δ_C = 138.7 (Ar), 138.6 (Ar), 138.4 (Ar), 138.2 (Ar), 138.1 (Ar), 137.8 (Ar), 135.9 (2C, Ar), 135.6 (2C, Ar), 135.1 (Ar), 133.9 (CH₂–CH=CH₂), 133.4 (Ar), 129.5 (2C, Ar), 128.6 (2C, Ar), 128.5 (2 C, Ar), 128.4 (4 C, Ar), 128.3 (2 C, Ar), 128.2 (2 C, Ar), 128.0 (3 C, Ar), 127.9 (2 C, Ar), 127.8 (6 C, Ar), 127.7 (2 C, Ar), 127.6 (3 C, Ar), 127.5 (2 C, Ar), 127.4 (4 C, Ar), 127.3 (Ar), 117.2 (CH₂–CH=CH₂), 99.3 (C-1''), 98.8 (C-1'), 97.9 (C-1'), 80.5, 80.0 (C-3, C-3''), 79.8 (C-3''), 75.2 (CH₂Ph), 75.1 (CH₂Ph), 74.8 (CH₂Ph), 74.7, 74.5 (C-4, C-4'), 74.2 (2 C, C-2', C-4''), 72.4 (C-5''), 72.0 (CH₂Ph), 71.9 (2 C, CH₂Ph), 71.7 (CH₂–CH=CH₂), 70.9 (2 C, C-5, C-5''), 68.4 (C-2), 68.1 (C-2''), 67.7 (CH₂O), 66.1, 65.9 (2 C, C-6, C-6'), 63.0 (C-6''), 51.4 (CH₂N₃), 29.4, 29.3, 29.1, 28.8 (4 C, OCH₂(CH₂)₆CH₂N₃), 26.9 (3 C, C(CH₃)₃), 26.7, 26.2 (2 C, OCH₂(CH₂)₆CH₂N₃), 19.4 (C(CH₃)₃). HRMS (ESI) calcd for (M + Na⁺) C₈₇H₁₀₅N₃O₁₆SiNa: 1498.7156. Found: 1498.7137.

8-Azidooctyl 2,3,4,6-Tetra-O-benzoyl-α-D-mannopyranosyl-(1→2)-3,4-di-O-benzyl-6-O-(tert-butyl)diphenylsilyl)-α-D-mannopyranosyl-(1→6)-2-O-allyl-3,4-di-O-benzyl-α-D-mannopyranosyl-(1→6)-[2,3,4,6-tetra-O-benzoyl-α-D-mannopyranosyl-(1→2)]-3,4-di-O-benzyl-α-D-mannopyranoside (36). A mixture of trichloroacetimidate **11b** (250 mg, 0.34 mmol), alcohol **35** (83 mg, 0.056 mmol), and 4 Å molecular sieves (25 mg) in Et₂O (780 μL) was stirred for 30 min at –15 °C under an argon atmosphere. Then, TMSOTf (325 μL of a 0.07 M solution in Et₂O) was added dropwise over 5 min. The reaction mixture was stirred for 2 h at –15 °C, and then the TMSOTf was quenched by the

addition of Et₃N. The solution was concentrated under vacuum and the resulting syrup was purified by column chromatography (9:1 hexane–acetone) to afford **36** (99 mg, 67%) as a syrup: R_f 0.41 (7:3 hexane–acetone); [α]_D + 1.3 (c = 0.08, CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ_H = 8.16 (d, J = 7.5 Hz, 2 H, Ar), 8.12 (d, J = 7.5 Hz, 2 H, Ar), 8.09 (d, J = 7.5 Hz, 2 H, Ar), 8.02 (d, J = 7.5 Hz, 2 H, Ar), 7.95 (d, J = 7.5 Hz, 2 H, Ar), 7.87 (d, J = 7.5 Hz, 2 H, Ar), 7.84–7.76 (m, 4 H, Ar), 7.71 (app t, J = 6.4 Hz, 4 H, Ar), 7.65–7.51 (m, 4 H, Ar), 7.51–7.05 (m, 56 H, Ar), 6.17 (t, 1 H, J = 10.2 Hz), 6.10 (t, 1 H, J = 9.7 Hz), 6.01–5.93 (m, 4 H), 5.88 (dddd, 1 H, J = 5.4, 6.2, 10.5, 17.2 Hz, CH₂–CH=CH₂), 5.40 (br s, 1 H), 5.26 (br s, 2 H), 5.21 (dd, 1 H, J = 1.2, 17.2 Hz, CH₂–CH=CH₂), 5.11 (br s, 1 H), 5.06 (d, 1 H, J = 11.1 Hz, CH₂Ph), 4.97–4.86 (m, 4 H), 4.81 (d, 1 H, J = 11.6 Hz, CH₂Ph), 4.78–4.66 (m, 8 H), 4.59–4.52 (m, 3 H), 4.50 (d, 1 H, J = 11.9 Hz, CH₂Ph), 4.49 (d, 1 H, J = 11.7 Hz, CH₂Ph), 4.39 (d, 1 H, J = 11.5 Hz, CH₂Ph), 4.30–4.21 (m, 4 H), 4.19 (br s, 1 H), 4.10 (br s, 1 H), 4.05–3.85 (m, 7 H), 3.83–3.54 (m, 7 H), 3.45 (d, 1 H, J = 11.2 Hz), 3.33 (dt, 1 H, J = 2 × 6.5, 9.5 Hz, OCH₂), 3.23 (t, 2 H, J = 7.0 Hz, CH₂N₃), 1.66–1.49 (m, 4 H, OCH₂(CH₂)₆CH₂N₃), 1.40–1.24 (m, 8 H, OCH₂(CH₂)₆CH₂N₃), 1.12 (s, 9 H, C(CH₃)₃); ¹³C NMR (151 MHz, CDCl₃) δ_C = 166.1 (2 C, C=O), 165.6 (C=O), 165.5 (C=O), 165.2 (C=O), 165.1 (C=O), 164.9 (2 C, C=O), 139.1 (Ar), 138.8 (Ar), 138.5 (Ar), 138.4 (Ar), 138.3 (Ar), 137.9 (Ar), 135.9 (3 C, Ar), 135.6 (3 C, Ar), 135.3 (Ar), 134.0 (Ar), 133.5 (CH₂–CH=CH₂), 133.4 (Ar), 133.3 (Ar), 133.2 (Ar), 133.1 (2 C, Ar), 133.0 (2 C, Ar), 132.9 (Ar), 130.0 (Ar), 129.9 (6 C, Ar), 129.8 (5 C, Ar), 129.7 (3 C, Ar), 129.6 (Ar), 129.5 (2 C, Ar), 129.4 (Ar), 129.1 (Ar), 129.0 (Ar), 128.9 (Ar), 128.5 (8 C, Ar), 128.4 (3 C, Ar), 128.3 (6 C, Ar), 128.2 (10 C, Ar), 128.1 (2 C, Ar), 128.0 (2 C, Ar), 127.9 (4 C, Ar), 127.7 (3 C, Ar), 127.6 (4 C, Ar), 127.5 (4 C, Ar), 127.3 (2 C, Ar), 127.2 (3 C, Ar), 127.1 (Ar), 116.8 (CH₂–CH=CH₂), 100.5, 99.7, 99.3, 99.0, 98.5, 79.9, 78.7, 78.5, 75.2 (2 C), 75.1, 74.8 (2 C), 74.5, 74.2, 73.9, 72.7, 72.4, 71.8, 71.7, 71.3, 71.2, 70.7, 70.3, 70.1, 70.0, 69.4, 69.2, 67.7 (CH₂O), 67.1, 66.9, 66.2, 65.8, 63.0 (3 C), 51.4 (CH₂N₃), 29.5, 29.4, 29.1, 28.8 (4 C, OCH₂(CH₂)₆CH₂N₃), 27.0 (3 C, C(CH₃)₃), 26.7, 26.2 (2 C, OCH₂(CH₂)₆CH₂N₃), 19.3 (C(CH₃)₃). HRMS (ESI) calcd for (M + Na⁺) C₁₅₅H₁₅₇N₃O₃₄SiNa: 2655.0315. Found: 2655.0310.

8-Azidooctyl 3,4-Di-O-benzyl-6-O-(tert-butyl)diphenylsilyl)-α-D-mannopyranosyl-(1→6)-2-O-allyl-3,4-di-O-benzyl-α-D-mannopyranosyl-(1→6)-3,4-di-O-benzyl-2-O-(4-methoxybenzyl)-α-D-mannopyranoside (37). To a solution of **9** (120 mg, 0.073 mmol) in Et₂O–CH₃OH (1:1, 500 μL) was added a solution of NaOCH₃ in CH₃OH (120 μL, 0.1M). The reaction mixture was stirred for 3 h, neutralized by the addition of Amberlyst IR-120 (H⁺) cation exchange resin, filtered, and concentrated to give a syrup that was purified by column chromatography (4:1 hexane–EtOAc) to afford **37** (113 mg, 97%) as a syrup. R_f 0.45 (8:2 hexane–EtOAc); [α]_D + 52.4 (c = 0.09, CH₂Cl₂); ¹H NMR (500 MHz, CDCl₃) δ_H = 7.75 (d, 2 H, J = 6.8 Hz, Ar), 7.69 (d, 2 H, J = 6.8 Hz, Ar), 7.46–7.09 (m, 38 H, Ar), 6.83 (d, 2 H, J = 8.4 Hz, PhOCH₃), 5.95 (dddd, 1 H, J = 5.4, 6.2, 10.5, 17.2 Hz, CH₂–CH=CH₂), 5.53 (dd, 1 H, J = 1.7, 2.9 Hz, H-2''), 5.34 (dd, 1 H, J = 1.4, 17.1 Hz, CH₂–CH=CH₂), 5.16 (dd, 1 H, J = 1.4, 10.4 Hz, CH₂–CH=CH₂), 5.07 (d, 1 H, J = 1.2 Hz, H-1'), 5.05 (d, 1 H, J = 1.7 Hz, H-1''), 4.94–4.87 (m, 3 H, CH₂Ph), 4.81 (d, 1 H, J = 1.1 Hz, H-1), 4.75–4.63 (m, 7 H, CH₂Ph), 4.61 (d, 1 H, J = 11.0 Hz, CH₂Ph), 4.56 (d, 1 H, J = 11.7 Hz, CH₂Ph), 4.48 (app t, 2 H, J = 10.6 Hz, CH₂Ph), 4.18 (br s, 1 H, H-2''), 4.15–4.10 (app d, 2 H, CH₂–CH=CH₂), 4.98 (t, 1 H, J = 9.5 Hz, H-4''), 4.01–3.86 (m, 7 H, H-4, H-4', H-3, H-3', H-3'', H-6'a, H-2'), 3.85–3.73 (m, 7 H, H-6a, H-6'a, PhOCH₃, H6''b, H-2), 3.72–3.58 (m, 6 H, H-6b, H-5, H-5', H-5'', H-6'b, CH₂O), 3.35 (dt, 1 H, J = 2 × 6.5, 9.5 Hz, OCH₂), 3.24 (t, 2 H, J = 6.9 Hz, CH₂N₃), 1.63–1.45 (m, 4 H, OCH₂(CH₂)₆CH₂N₃), 1.42–1.23 (m, 8 H, OCH₂(CH₂)₆CH₂N₃), 1.06 (s, 9 H, C(CH₃)₃); ¹³C NMR (126 MHz, CDCl₃) δ_C = 159.3 (Ar), 138.7 (Ar), 138.6 (Ar), 138.5 (Ar), 138.3 (Ar), 138.0 (Ar), 135.9 (2C, Ar), 135.7 (2C, Ar), 135.3 (Ar), 133.9 (CH₂–CH=CH₂), 133.4 (Ar), 130.4 (Ar), 129.5 (4C, Ar), 128.5 (2C, Ar), 128.4 (4 C, Ar), 128.3 (2 C, Ar), 128.2 (4 C, Ar), 128.1 (2 C, Ar), 128.0 (2 C, Ar), 127.8 (Ar), 127.7 (5 C, Ar), 127.6 (8 C, Ar), 127.5 (1 C, Ar), 127.4 (3

C, Ar), 127.3 (Ar), 116.9 (CH₂-CH=CH₂), 113.8 (2 C, Ar), 99.8 (C-1'), 98.1 (C-1', 98.0 (C-1), 80.5 (C-3), 79.8 (C-3''), 79.4 (C-3'), 75.1 (CH₂Ph), 75.0 (CH₂Ph), 74.9 (CH₂Ph), 74.8 (C-2'), 74.6 (C-2), 74.5, 74.3 (C-4, C-4'), 74.1 (C-4''), 72.5 (CH₂Ph), 72.3 (C-5'), 72.1 (CH₂Ph), 71.8 (CH₂Ph), 71.7 (2 C, C-5, C-5'), 71.6 (CH₂Ph), 71.5 (CH₂-CH=CH₂), 68.1 (C-2''), 67.6 (CH₂O), 66.0 (2 C, C-6, C-6'), 62.9 (C-6''), 55.2 (PhOCH₃), 51.4 (CH₂N₃), 29.4, 29.3, 29.1, 28.8 (4 C, OCH₂(CH₂)₆CH₂N₃), 26.8 (3 C, C(CH₃)₃), 26.7, 26.1 (2 C, OCH₂(CH₂)₆CH₂N₃), 19.3 (C(CH₃)₃). HRMS (ESI) calcd for (M + Na⁺) C₉₅H₁₁₃N₃O₁₇SiNa: 1618.7731. Found: 1618.7719.

8-Azidoctyl 2,3-Di-O-benzoyl-6-O-(tert-butylidiphenylsilyl)- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 2)-3,4-di-O-benzyl-6-O-(tert-butylidiphenylsilyl)- α -D-mannopyranosyl-(1 \rightarrow 6)-2-O-allyl-3,4-di-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6)-3,4-di-O-benzyl-2-O-(4-methoxybenzyl)- α -D-mannopyranoside (38).

The synthesis of **38** was achieved following the procedure described for the preparation of **25**, using alcohol **37** (100 mg, 0.071 mmol), thioglycoside **12** (147 mg, 0.071 mmol), and 4 Å molecular sieves (23 mg) in CH₂Cl₂ (2.4 mL) in the presence of NIS (19 mg, 0.084 mmol) and AgOTf (4 mg, 0.016 mmol). The crude residue was purified by column chromatography (9:1 \rightarrow 4:1, hexane-acetone) to yield **38** (164 mg, 74%) as a syrup. *R*_f 0.74 (3:2 hexane-acetone); [α]_D + 25.9 (*c* = 0.17, CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ _H = 8.10–7.86 (m, 20 H, Ar), 7.79–7.65 (m, 8 H, Ar), 7.62–7.04 (m, 74 H, Ar), 6.83 (d, 1 H, *J* = 8.4 Hz, Ph-OMe), 5.93 (dddd, 1 H, *J* = 5.4, 6.2, 10.5, 17.2 Hz, CH₂-CH=CH₂), 5.77 (d, 1 H, *J* = 4.6 Hz), 5.75–5.65 (m, 9 H), 5.61 (br s, 1 H), 5.47–5.39 (m, 4 H, 4 \times H-1), 5.31 (dd, 1 H, *J* = 1.5, 17.2 Hz, CH₂-CH=CH₂), 5.13 (dd, 1 H, *J* = 1.5, 10.5 Hz, CH₂-CH=CH₂), 5.05 (br s, 2 H, 2 \times H-1), 4.98–4.88 (m, 3 H, CH₂Ph), 4.82 (br s, 1 H, H-1), 4.76 (d, 1 H, *J* = 11.7 Hz, CH₂Ph), 4.72–4.42 (m, 15 H), 4.38 (br s, 1 H), 4.27–4.10 (m, 7 H, CH₂-CH=CH₂), 4.05–3.57 (m, 25 H), 3.52 (app d, 1 H, *J* = 10.6 Hz, H-6), 3.34 (dt, 1 H, *J* = 2 \times 6.5, 9.5 Hz, OCH₂), 3.24 (t, 2 H, *J* = 6.9 Hz, CH₂N₃), 1.67–1.46 (m, 4 H, OCH₂(CH₂)₆CH₂N₃), 1.41–1.22 (m, 8 H, OCH₂(CH₂)₆CH₂N₃), 1.06 (br s, 18 H, 2 \times C(CH₃)₃); ¹³C NMR (126 MHz, CDCl₃) δ _C = 165.7 (C=O), 165.6 (3 C, C=O), 165.5 (C=O), 165.2 (C=O), 165.1 (2 C, C=O), 165.0 (2 C, C=O), 159.3 (Ar), 139.0 (Ar), 138.8 (Ar), 138.6 (Ar), 138.5 (Ar), 138.3 (Ar), 138.2 (Ar), 136.0 (2 C, Ar), 135.7 (6 C, Ar), 135.6 (2 C, Ar), 135.2 (Ar), 134.1 (Ar), 133.6 (CH₂-CH=CH₂), 133.3 (2 C, Ar), 133.1 (4 C, Ar), 133.0 (Ar), 130.4 (Ar), 130.0 (3 C, Ar), 129.9 (10 C, Ar), 129.8 (7 C, Ar), 129.7 (2 C, Ar), 129.5 (3 C, Ar), 129.4 (Ar), 129.3 (Ar), 129.2 (2 C, Ar), 129.1 (2 C, Ar), 128.5 (5 C, Ar), 128.4 (13 C, Ar), 128.3 (11 C, Ar), 128.2 (4 C, Ar), 128.1 (5 C, Ar), 128.0 (2 C, Ar), 127.9 (2 C, Ar), 127.7 (10 C, Ar), 127.6 (8 C, Ar), 127.5 (2 C, Ar), 127.4 (Ar), 127.2 (2 C, Ar), 127.1 (Ar), 117.0 (CH₂-CH=CH₂), 113.7 (2 C, Ar), 106.3 (C-1), 106.0 (3 \times C-1), 105.9 (C-1), 99.6 (C-1), 98.4 (C-1), 97.9 (C-1), 83.2, 82.3, 82.1 (6 C), 81.9, 81.5 (3 C), 80.5, 79.9, 79.6, 77.2, 75.1, 75.0, 74.7, 74.6 (2 C), 74.5 (2 C), 74.3, 73.1, 72.5, 72.4, 72.1, 71.9, 71.7, 71.6, 71.4, 71.3, 67.5 (2 C), 66.1, 66.0, 65.9, 65.8 (2 C), 65.5, 63.4 (2 C), 63.1, 55.2 (PhOCH₃), 51.4 (CH₂N₃), 29.4, 29.3, 29.1, 28.8 (4 C, OCH₂(CH₂)₆CH₂N₃), 26.8 (6 C, 2 \times C(CH₃)₃), 26.7, 26.2 (2 C, OCH₂(CH₂)₆CH₂N₃), 19.3 (2 C, 2 \times C(CH₃)₃). HRMS (ESI) calcd for (M + 2 Na⁺) C₂₀₆H₂₁₁N₃O₄₇Si₂Na₂: 1790.1768. Found: 1790.1774.

8-Azidoctyl 2,3-Di-O-benzoyl-6-O-(tert-butylidiphenylsilyl)- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 2)-3,4-di-O-benzyl-6-O-(tert-butylidiphenylsilyl)- α -D-mannopyranosyl-(1 \rightarrow 6)-2-O-allyl-3,4-di-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6)-3,4-di-O-benzyl-2-O-(4-methoxybenzyl)- α -D-mannopyranoside (39). The synthesis of **39** was achieved following the procedure described for the synthesis of **30**, starting from the octasaccharide **38** (52 mg, 0.015 mmol) using CAN (16 mg, 0.029 mmol) in CH₃CN-H₂O (10:1, 1.1 mL). The crude residue was purified by column chromatography (99:1, CH₂Cl₂-acetone) to yield **39** (34 mg, 68%) as a syrup. *R*_f 0.53 (3:2 hexane-

EtOAc); [α]_D + 24.4 (*c* = 0.11, CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ _H = 8.05–7.81 (m, 20 H, Ar), 7.74–7.61 (m, 8 H, Ar), 7.58–7.04 (m, 72 H, Ar), 5.90 (dddd, 1 H, *J* = 5.4, 6.2, 10.5, 17.2 Hz, CH₂-CH=CH₂), 5.73 (d, 1 H, *J* = 4.9 Hz), 5.70–5.60 (m, 9 H), 5.56 (br s, 1 H), 5.43–5.33 (m, 4 H, 4 \times H-1), 5.28 (dd, 1 H, *J* = 1.5, 17.2 Hz, CH₂-CH=CH₂), 5.12 (dd, 1 H, *J* = 1.5, 10.5 Hz, CH₂-CH=CH₂), 5.01 (d, 1 H, *J* = 1.2 Hz, H-1), 4.97 (d, 1 H, *J* = 1.5 Hz, H-1), 4.92–4.81 (m, 4 H, H-1, 3 \times CH₂Ph), 4.74–4.41 (m, 15 H), 4.34 (br s, 1 H), 4.22–4.06 (m, 8 H), 4.02 (br s, 1 H), 4.00–3.78 (m, 13 H), 3.76–3.57 (m, 6 H), 3.50 (app d, 1 H, *J* = 10.6 Hz, H-6), 3.35 (dt, 1 H, *J* = 2 \times 6.5, 9.5 Hz, OCH₂), 3.21 (t, 2 H, *J* = 6.9 Hz, CH₂N₃), 2.37 (d, 1 H, *J* = 2.2 Hz, OH), 1.62–1.47 (m, 4 H, OCH₂(CH₂)₆CH₂N₃), 1.37–1.21 (m, 8 H, OCH₂(CH₂)₆CH₂N₃), 1.01 (br s, 18 H, 2 \times C(CH₃)₃); ¹³C NMR (126 MHz, CDCl₃) δ _C = 165.6 (4 C, C=O), 165.5 (C=O), 165.2 (C=O), 165.1 (2 C, C=O), 165.0 (2 C, C=O), 138.9 (Ar), 138.8 (Ar), 138.3 (3 C, Ar), 137.9 (Ar), 136.0 (2 C, Ar), 135.7 (6 C, Ar), 135.6 (2 C, Ar), 135.2 (Ar), 134.1 (Ar), 133.5 (CH₂-CH=CH₂), 133.3 (4 C, Ar), 133.2 (2 C, Ar), 133.1 (4 C, Ar), 133.0 (Ar), 130.0 (3 C, Ar), 129.9 (5 C, Ar), 129.8 (10 C, Ar), 129.6 (2 C, Ar), 129.4 (2 C, Ar), 129.3 (2 C, Ar), 129.2 (2 C, Ar), 129.1 (3 C, Ar), 128.6 (2 C, Ar), 128.5 (5 C, Ar), 128.4 (9 C, Ar), 128.3 (7 C, Ar), 128.2 (8 C, Ar), 128.1 (4 C, Ar), 128.0 (Ar), 127.9 (6 C, Ar), 127.7 (9 C, Ar), 127.6 (5 C, Ar), 127.5 (2 C, Ar), 127.3 (Ar), 127.2 (2 C, Ar), 127.1 (Ar), 117.3 (CH₂-CH=CH₂), 106.4 (C-1), 106.0 (2 \times C-1), 105.9 (2 \times C-1), 99.5 (C-1), 99.0 (C-1), 98.4 (C-1), 83.2 (2 C), 82.2, 82.1 (4 C), 81.9, 81.5 (4 C), 80.5, 79.9, 79.8, 75.1, 75.0, 74.7, 74.6, 74.5, 74.4, 74.0, 73.1, 72.5, 71.9 (4 C), 71.8, 71.6, 71.4, 70.9, 68.3, 67.6, 66.1, 66.0, 65.9, 65.8, 65.7, 65.5, 63.4 (2 C), 63.1, 51.4 (CH₂N₃), 29.4, 29.3, 29.1, 28.8 (4 C, OCH₂(CH₂)₆CH₂N₃), 26.8 (6 C, 2 \times C(CH₃)₃), 26.7, 26.1 (2 C, OCH₂(CH₂)₆CH₂N₃), 19.3 (2 C, 2 \times C(CH₃)₃). HRMS (ESI) calcd for (M + 2 Na⁺) C₁₉₈H₂₀₃N₃O₄₆Si₂Na₂: 1730.1480. Found: 1730.1507.

8-Azidoctyl 2,3-Di-O-benzoyl-6-O-(tert-butylidiphenylsilyl)- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 5)-2,3-di-O-benzoyl- α -D-arabinofuranosyl-(1 \rightarrow 2)-3,4-di-O-benzyl-6-O-(tert-butylidiphenylsilyl)- α -D-mannopyranosyl-(1 \rightarrow 6)-2-O-allyl-3,4-di-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6)-[2,3,4,6-tetra-O-benzoyl- α -D-mannopyranosyl-(1 \rightarrow 2)]-3,4-di-O-benzyl- α -D-mannopyranoside (40). The synthesis of **40** was achieved following the procedure described for the preparation of **36**, using alcohol **39** (26 mg, 0.0076 mmol), trichloroacetimidate **11b** (23 mg, 0.031 mmol), and 4 Å molecular sieves (6 mg) in CH₂Cl₂ (400 μ L) in the presence of TMSOTf (22 μ L of a 0.07 M solution in CH₂Cl₂). The crude residue was purified by gel filtration chromatography (Sephadex, LH-20) with (1:1, CH₂Cl₂-CH₃OH) as the eluent, followed by column chromatography (99:1, CH₂Cl₂-acetone) to yield **40** (25 mg, 82%) as a syrup. *R*_f 0.73 (99:1, CH₂Cl₂-acetone); [α]_D + 6.5 (*c* = 0.60, CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ _H = 8.13–7.81 (m, 25 H, Ar), 7.75–7.52 (m, 12 H, Ar), 7.51–7.02 (m, 83 H, Ar), 6.08 (t, 1 H, *J* = 9.9 Hz, H-4''), 5.97–5.87 (m, 3 H, H-2'', H-3'', CH₂-CH=CH₂), 5.74 (d, 1 H, *J* = 4.7 Hz), 5.71–5.59 (m, 9 H), 5.55 (br s, 1 H), 5.43–5.33 (m, 5 H, 4 \times H-1, H-1''), 5.26 (dd, 1 H, *J* = 1.5, 17.2 Hz, CH₂-CH=CH₂), 5.07–5.03 (m, 2 H, H-1, CH₂-CH=CH₂), 5.01 (d, 1 H, *J* = 1.2 Hz, H-1), 4.97 (d, 1 H, *J* = 1.5 Hz, H-1), 4.93–4.83 (m, 4 H, H-1, 3 \times CH₂Ph), 4.78 (d, 1 H, *J* = 11.5 Hz, CH₂Ph), 4.74–4.42 (m, 15 H), 4.39 (d, 1 H, *J* = 11.5 Hz, CH₂Ph), 4.35 (br s, 1 H), 4.30–4.06 (m, 9 H), 4.03–3.79 (m, 13 H), 3.77–3.52 (m, 6 H), 3.46 (app d, 1 H, *J* = 10.9 Hz, H-6), 3.29 (dt, 1 H, *J* = 2 \times 6.5, 9.5 Hz, OCH₂), 3.20 (t, 2 H, *J* = 6.9 Hz, CH₂N₃), 1.58–1.41 (m, 4 H, OCH₂(CH₂)₆CH₂N₃), 1.37–1.19 (m, 8 H, OCH₂(CH₂)₆CH₂N₃), 1.01 (br s, 18 H, 2 \times C(CH₃)₃); ¹³C NMR (126 MHz, CDCl₃) δ _C = 166.1 (C=O), 165.7 (C=O), 165.6 (3 C, C=O), 165.5 (2 C, C=O), 165.3 (C=O), 165.2 (C=O), 165.1 (2 C, C=O), 165.0 (2 C, C=O), 164.9 (C=O), 139.0 (Ar), 138.8 (Ar), 138.4 (Ar), 138.3 (Ar), 138.2 (Ar), 137.9 (Ar), 135.9 (2 C, Ar), 135.7 (6 C, Ar), 135.6 (2 C, Ar), 135.3 (Ar), 134.1 (Ar), 133.6 (Ar), 133.5 (CH₂-CH=CH₂), 133.3 (4 C, Ar), 133.2 (2 C, Ar), 133.1 (4 C, Ar), 133.0 (Ar), 130.0 (4 C, Ar), 129.9 (9 C, Ar), 129.8 (15 C, Ar), 129.7 (2 C, Ar), 129.6 (3 C, Ar), 129.4 (4 C, Ar), 129.3 (2 C, Ar),

129.2 (2 C, Ar), 129.1 (4 C, Ar), 128.9 (Ar), 128.5 (8 C, Ar), 128.4 (6 C, Ar), 128.3 (11 C, Ar), 128.2 (11 C, Ar), 128.1 (3 C, Ar), 128.0 (4 C, Ar), 127.9 (2 C, Ar), 127.7 (10 C, Ar), 127.6 (4 C, Ar), 127.5 (4 C, Ar), 127.4 (Ar), 127.3 (Ar), 127.2 (Ar), 127.0 (2 C, Ar), 116.8 (CH₂-CH=CH₂), 106.3 (C-1), 106.0 (2 × C-1), 105.9 (2 × C-1), 99.6 (C-1), 99.3 (C-1), 99.0 (C-1), 98.5 (C-1), 83.2 (2 C), 82.2, 82.1 (5 C), 82.0, 81.5 (4 C), 80.1, 79.8, 75.1 (2 C), 75.0, 74.7 (2 C), 74.4 (2 C), 74.1, 73.0, 72.7, 72.3 (2 C), 71.6, 71.4 (3 C), 71.2, 70.1, 70.0, 69.4, 67.7, 67.0, 65.9 (2 C), 65.8 (2 C), 65.7, 65.5, 63.4 (2 C), 63.0 (2 C), 59.6 (2 C), 51.4 (CH₂N₃), 29.5, 29.4, 29.1, 28.8 (4 C, OCH₂(CH₂)₆CH₂N₃), 26.7, 26.2 (6 C, 2 × C(CH₃)₃), 19.3 (2 C, 2 × (C(CH₃)₃)); HRMS (ESI) calcd for (M + 2 Na⁺) C₂₃₂H₂₂₉N₃O₃₅Si₂Na₂: 2019.2269. Found: 2019.2287.

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.5b02083.

NMR spectra for all new compounds (PDF)

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Notes

The authors declare no competing financial interest.

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